ORIGINAL PAPER



Dietary variation in a family group of the woolly false vampire Bat (*Chrotopterus auritus*, Phyllostomidae) at the north of its distribution

Silvia Mónica Cerecedo-Jiménez¹ · Pedro Adrián Aguilar-Rodríguez² · M. Cristina MacSwiney G.³ · Martín Alarcón Montano² · Viridiana Vega-Badillo⁴ · Nallely Verónica Rodríguez Santiago⁵ · José Luis Aguilar López⁶

Received: 23 December 2024 / Accepted: 19 June 2025

© The Author(s), under exclusive licence to Mammal Research Institute Polish Academy of Sciences 2025

Abstract

Carnivorous bats are species that feed on terrestrial vertebrates year-round or seasonally, a food habit observed in four families. Within Phyllostomidae, *Chrotopterus auritus* is one of the largest Neotropical bat species with a diet mainly consisting of small vertebrates. Few studies have detailed the diet of carnivorous bats throughout the year. Here, we recorded the diet of a family group of *C. auritus* weekly during a year in the northernmost location of its distribution, in the Los Tuxtlas region (Mexico). Dietary items were collected from guano in a cave roost. The food items were separated and classified [mammal, bird, insects and unidentified soft-tissue], dried and weighed, and identified to the lowest possible taxonomic level based on morphological characteristics. We recorded each item as a single incidence to calculate frequency and abundance. We estimated prey category diversity using morphospecies richness. Prey morphospecies diversity did not differ between the dry and windy seasons, but both had higher diversity than the rainy season. Mammal remains had the highest biomass (62.41% of sample weight) and were consumed especially during the rainy season, as well as bird remains. Insects peaked in the windy season but were found all year. We found that this group of *C. auritus* preys on vertebrates during its reproductive season and pup-rearing period, while relying on insects as a year-round food source.

Keywords Carnivory · Insects · Birds · Small mammals · Los Tuxtlas

Communicated by: Facundo Luna

Pedro Adrián Aguilar-Rodríguez pedroaguilarr@gmail.com

Silvia Mónica Cerecedo-Jiménez monicacerecedo1@gmail.com

M. Cristina MacSwiney G. cmacswiney@uv.mx

Martín Alarcón Montano alarconmom18@gmail.com

Viridiana Vega-Badillo viridiana.vega@inecol.mx

Nallely Verónica Rodríguez Santiago nalrodriguez@uv.mx

José Luis Aguilar López jose.aguilarlopez@ecosur.mx

Published online: 17 July 2025

Facultad de Biología, Universidad Veracruzana, Zona Universitaria, 91090 Xalapa, Veracruz, México

- Instituto de Investigaciones Forestales, Universidad Veracruzana, Parque Ecológico El Haya, S/N, 91070 Xalapa, Veracruz, México
- ³ Centro de Investigaciones Tropicales, Universidad Veracruzana, Calle José María Morelos y Pavón 44, 91000 Xalapa, Veracruz, México
- Colección Entomológica IEXA "Dr. Miguel Ángel Morón Ríos, Instituto de Ecología, A. C., Carretera Antigua a Coatepec 351, 91073 Xalapa, Veracruz, México
- Colección de Mamíferos (IIB-UV), Instituto de Investigaciones Biológicas, Dr. Luis Castelazo Ayala, S/N, 91190 Xalapa, Veracruz, México
- Departamento de Conservación de la Biodiversidad, El Colegio de la Frontera Sur, Carretera Panamericana y Periférico Sur, S/N, 29290 San Cristóbal de las Casas, Chiapas, México

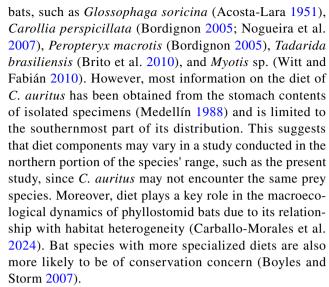


Introduction

In Chiroptera, carnivory is a feeding habit in which a bat species consumes terrestrial vertebrates year-round or seasonally as the main component of its diet (Wilson 1979; Hill and Smith 1984; Gual-Suárez and Medellín 2021). The families in which carnivorous habits are observed include Phyllostomidae, Megadermatidae, Nycteridae, and Vespertilionidae (Gardner 1977; Wilson 1979; Hill and Smith 1984). Within the Phyllostomidae family, five Mexican species (out of 61 species in the family for the country; Simmons and Cirranello 2025) have vertebrates as significant components of their diets ('75% of their diet): the Greater False Vampire Bat (Vampyrum spectrum), the Woolly False Vampire Bat, (Chrotopterus auritus), the Keenan's Hairy-nosed Bat (Gardnerycteris keenani), the Cozumelan Golden Bat (Mimon cozumelae), and Coffin's Bat (Trachops coffini) (Gual-Suárez and Medellín 2021).

Chrotopterus auritus, is one of the largest bat species in the Neotropics, with a forearm between 77–83 mm. In comparison, Vampyrum spectrum's is between 98–110 mm and Phyllostomus hastatus 80-93 mm (Navarro and Wilson 1982; Santos et al. 2003). Chrotopterus auritus ranges from southern Mexico to northern Argentina and Paraguay (Barquez et al. 2015). This species typically occurs in small groups composed of a reproductive couple and one or two pups (Medellín 1989; Linares 1998; Ochoa et al. 2005). It occupies a variety of roosting sites, including caves, termite nests, hollow trees, dead trunks and abandoned human structures (Gual-Suárez and Medellín 2021), with roosts reaching up to 30 °C and high humidity levels (Gual-Suárez et al. 2025). The maximum recorded flight distance is 2.06 km, with a total of up to 5.3 km traveled per night (Vleut et al. 2019). Individuals spend approximately five hours outside the roost each night (Vleut et al. 2019). Studies show that its diet primarily consists of small mammals, representing more than 70% of the biomass consumed, and may even include other bat species (Bonato et al. 2004; Nogueira et al. 2007), with no significant dietary differences between sexes (Bonato et al. 2004), it is also categorized as a gleaning animalivorous species (de Oliveira et al. 2017).

Among the species known to be consumed by *C. auritus* are various vertebrates, including rodents (Oryzomyinae; *Ototylomys phyllotis, Heteromys goldmani, Reithrodontomys mexicanus, Peromyscus oaxacensis, P. guatemalensis*, and *Nyctomys sumichrasti*), mouse opossums (*Marmosa* sp.), shrews (*Sorex* spp.), doves (*Columbigallina talpacoti*), *Knipolegus cabanisi*; geckos (*Thecadactylus rapicaudus*), anurans (possibly Hylidae) (Medellín 1988). There are also reports of *C. auritus* preying on other



Although carnivorous bats consume both vertebrates and invertebrates (Wilson 1979; Hill and Smith 1984), few studies have detailed their diets, including how the proportions of different animal taxa vary throughout the year. This study presents, for the first time, the annual variation in the diet of C. auritus, which is an endangered species in Mexico (SEMARNAT 2010), in the northernmost portion of its range, specifically in the Los Tuxtlas region of Veracruz, Mexico. We monitored a small group of C. auritus and documented their diet over the course of a year. We expect that the diet would consist mainly of vertebrates and would be more diverse during the rainy season or during the species' reproductive period. By identifying the food items consumed by this species, we gain insights into its ecological niche (Carvalho et al. 2020) and its vulnerability to habitat transformation, especially in a context where prey for carnivorous bats may vary both geographically and seasonally.

Materials and methods

Study area

Samples were collected in Ejido Adolfo Ruiz Cortines (18° 32′ 40.04″ N, 95° 09′ 2.20″ W, 1082 m), located within the Los Tuxtlas Biosphere Reserve, on the southeastern slope of the San Martín Tuxtla volcano (municipality of San Andrés, Tuxtla) from January 2019 to December 2019. The mean annual temperature in this area is 18 °C, and the mean annual precipitation is approximately 4000 mm (Soto 2004). The study area experiences three seasons; rainy season (June to October); windy season (November to February); and dry season (March to May; Soto 2004; Nogueira et al. 2007). The original vegetation was montane cloud



forest, now largely reduced to remnants surrounded by a matrix of secondary vegetation, crops, and cattle pastures (Castillo-Campos and Laborde 2004).

Prey remains collection

Dietary items were collected from guano and prey remains left in a cave roost occupied by a group of specimens of Chrotopterus auritus, consisting of a reproductive pair and one or two juveniles, which is typical for this species (Araujo and Machado 2012). No more individuals of this species were registered in any inspection of the cave (or in nearby caves), even during our mist-netting sampling at the three entrances of the cave prior to this study (more than 20 sampling nights over three years before this study). Other bats inhabiting this cave include Mormoopidae (Mormoops megalophylla, Pteronotus mesoamericanus, Pteronotus fulvus), Phyllostomidae; (Artibeus jamaicensis, Artibeus toltecus, Glossophaga mutica, Desmodus rotundus), Vespertilionidae (Myotis extremus and Myotis elegans), none of which are carnivorous bat species. Since vertebrate-eating bats tend to leave prey remains (Medellín 1988; Jordan 2005; Felix et al. 2013), we conducted preliminary daytime inspections to locate individual roosting sites within the cave where the C. auritus group rested. We identified areas containing food remains and guano directly below perching bats-hereafter referred to as "feeding sites". Over the months leading up to sample collection, we established four feeding sites within the cave, used consistently by C. auritus individuals during feeding, where prey remains accumulated beneath them.

We inspected these areas weekly for prey remains and guano containing partially consumed items. At each feeding site, we cleared a 1 m^2 section of the substrate and placed a plastic sheet over it to collect additional material (Whitaker et al. 1981, 2009; Martínez-Fonseca et al. 2022). Two people examined the plastic sheets each week, manually collecting remains and guano using tweezers. Samples were stored in resealable plastic bags containing 70% ethanol (one bag per week) and refrigerated until further analysis (Gardner 1977; McAney et al. 1991; Martínez-Fonseca et al. 2022).

We also recorded field observations when bats were seen perching above the feeding sites, noting the number of individuals, their relative size, and fur coloration to identify juveniles or pups. Sample collection was completed in under five minutes per site using dim red light, minimizing disturbance to roosting bats. Unless directly illuminated by a headlamp, bats rarely responded to the researchers' presence. The most frequent behavior observed was adult individuals "hugging" the juveniles.

Laboratory work

In the laboratory, each bag was cleaned and processed. Ethanol was poured out, and the contents were transferred to Petri dishes and rinsed with distilled water to remove any debris. Food remains were manually separated using tweezers and classified as mammal remains (bones and hair), bird remains (hollow bones and feathers), arthropod exoskeleton parts (Bonato et al. 2004) or unidentified soft tissues. The remains, including soft tissues, were placed in waxed paper bags to dry with silica gel desiccant (Hansson 1970), then weighed using a digital scale (Ohaus Mod. PA214). Samples containing soft tissue were also cleaned with distilled water, weighed, and preserved in 70% ethanol (Whitaker et al. 2009). Due to the fragmented condition of the remains, most items were identified only to morphospecies, limiting taxonomic resolution.

Putative mammal bones, skull fragments, and hair were examined using a stereoscopic microscope and taxonomic guides (Hall and Kelson 1959; Altenbach 1979; Huckaby 1980; Reid 2009; Gardner 2008; Ceballos and Oliva 2002). We compared them with specimens from the Mammal Collection at Universidad Veracruzana to determine the lowest possible taxonomic category. Cusp morphology was measured on the upper and lower teeth, jaws, and palates, and compared with corresponding structures in local mammal specimens. This process was also used to identify other bones, including bat forearms, humeri (evaluating humeral head and epicondyle shape and size), sternum, ribs and vertebrae. Leg and tail remains (with soft tissue, nails and hair attached) were measured, photographed, and compared with taxidermized specimens from the collection, then verified with taxonomic keys. Rodent taxonomy followed Álvarez et al. (2015), while Choate (1973) and Woodman (2018) were used for shrews and bats, with names validated by Díaz et al. (2021) and Da Silva Fonseca et al. (2024). Additionally, several bone clusters were found in fur clumps, suggesting each clump represents an individual. Unclassifiable tissue remains were cleaned, photographed, and stored in vials with 70% alcohol.

Feathers were identified by cross-referencing with the local bird species list, considering features such as the rachis size and shape, tip shape, and plumage color, following Slud (1964), Howell and Webb (1995), Baptista et al. (1997), Johnsgard (1997), Schodde and Manson (1997), Peterson and Chalif (1989), and Howell (2002). Identification guides, including The Taxonomy of Avibase (1766), Cory (1918), and Bernis et al. (1998), were used for scientific naming. Some feathers were identified as juvenile due to their clustering, with bones found within the clumps, suggesting they may belong to the same individual.



Chitinous insect remains (elytra, legs, etc.) were mounted on a 1 cm² piece of sulfated cardboard using entomological glue and sorted by morphotype (Whitaker et al. 2009). Identification was performed using morphological characteristics and taxonomic keys (White 1983; Arnett et al. 2002; Leschen et al. 2010).

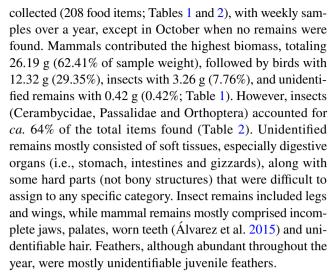
Data analysis

Weekly samples were weighed and analyzed per season to identify which category had the highest biomass (Kunz et al. 1995; Whitaker et al. 2009; Borloti et al. 2019). In addition, we recorded each observed item (feathers, bones, wings, nails, etc.) as a single occurrence in each category: mammals, birds, insects (i.e., two different feathers were counted as 2 for the "bird" category). These counts were summed per season to determine seasonal abundance (Jordan 2005; Whitaker et al. 2009). We also calculated relative frequency, defined as the number of items of a given taxon divided by the total number of items. Both biomass and frequency data were presented due to the nature of the samples-fragmented remains mixed with fur and feathers- and the inability to separate individual prey items. Frequency-based proportions are indicative rather than precise representations of diet (Jordan 2005).

To compare prey category diversity across seasons, we estimated diversity using Hill numbers: 0D (species richness), 1D (effective number of common prey categories), and 2D (effective number of abundant prey categories) across the three seasons studied (dry, rainy, and windy). These calculations were based on proportional prey category abundances. First, to guarantee a fair comparison among data sets, we assessed sample coverage for each season using the Chao non-parametric estimator (Chao et al. 2014). If 100% sample coverage was not achieved, even with doubled sample extrapolation, we applied standardized sample coverage (Chao et al. 2014). We then estimated 84% confidence intervals from 1000 bootstrap iterations, as they better approximate a 0.05 significance test than 95% intervals (MacGregor and Payton 2013). All calculations were conducted in R using the iNEXT package (Hsieh et al. 2020; R Development Core Team 2022).

Results

The composition of the family group of *Chrotopterus auritus* changed throughout the year. Most of the year, a pair of reproductive individuals cohabited with a subadult. However, by the end of the dry season, we observed the presence of a younger pup, bringing the group to four individuals (two reproductive adults, the previous year's subadult, and the pup; Fig. 1, Video S1). A total of 44 samples were



Mammalian remains identified belonged to the families Soricidae (e.g., Nelson's Small-eared Shrew [Cryptotis nelsoni]), Cricetidae (Nyctomys sp. [Vesper Rat], Oryzomys alfaroi [Alfaro's Rice Rat], Peromyscus mexicanus [Mexican Deer Mouse], and P. leucopus [White-footed Mouse]), Phyllostomidae (Artibeus jamaicensis [Jamaican Fruit Bat], Desmodus rotundus [Common Vampire Bat]), and the Mormoopidae family (leaf-chinned bats), which were observed only in September (Figs. 2 and 3; Table 2). Bird feathers from the families Columbidae (pigeons and doves) (determined as Columbina sp.), Thraupidae (tanagers) (Cyanerpes cyaneus) and Trochilidae (hummingbirds) were also identified (Fig. 4), the former being found across seasons (Table 2). Insect remains included two orders, Coleoptera (beetles) (five families identified) and Orthoptera (no family identification). Orthoptera (crickets and grasshoppers; Fig. 5) were found in seven months, while Coleoptera appeared in eight months (Table 2). Cerambycidae, Passalidae and Orthoptera were present throughout all seasons. The "Other" category contained unclassifiable samples, mostly soft tissues (primarily viscera).

Mammals were consumed throughout the year, with the highest occurrence of mammalian remains in September, coinciding with the rainy season and a decrease in March (Fig. 1). Bird remains were more common in the rainy season, while insect remains peaked in the windy season. Notably, insect consumption spiked in December during the windy season, which overlaps with the mating season (January-August), while insect consumption was absent in August and November (Fig. 1).

Sample coverage during the dry season did not reach 100%, even with doubled sample size extrapolation, so we standardized coverage to 79% across seasons. Prey category richness (0D) was similar between the dry and windy seasons, but both were higher than in the rainy season (Fig. 6). Effective diversity (1D) and dominance (2D) were higher in the windy season compared to the rainy season,



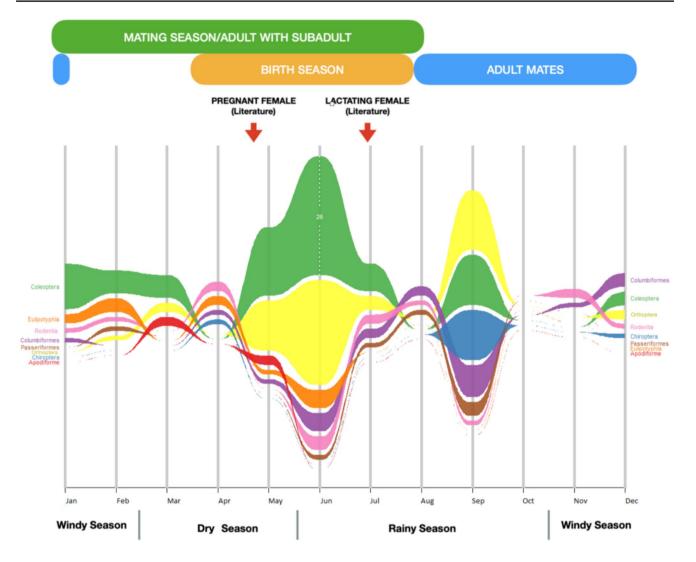


Fig. 1 Alluvial diagram showing the occurrence of prey items in the diet of a group of *Chrotopterus auritus* through a year. Each color represents an order and its occurrence in the diet (each time a prey item in such category was found), by month. The line thickness

increases with higher incidence. The figure also depicts the three climatic seasons and the life stages of *C. auritus*, based on our observations and consulted literature (e.g., Wilson 1979; Álvarez et al. 2015)

Table 1 Mean biomass (in g) and error data of the four categories of remains consumed by a family of *Chrotopterus auritus*, categorized by climatic season throughout the year

Item Category	Windy Season	Dry Season	Rainy Season
Mammals	0.40 ± 0.28	0.72 ± 0.51	5.62 ± 5.95
Birds	0.13 ± 0.08	0.42 ± 0.34	2.63 ± 1.97
Insects	0.27 ± 0.32	0.10 ± 0.08	0.47 ± 0.45
Others*	0.02 ± 0.02	0.01 ± 0.01	0.02 ± 0.04

but not significantly different from the dry season. Overall, the diversity analysis indicates that the rainy season has lower values than the dry and windy seasons (Fig. 6).

Discussion

Prey remains found in the refuge of *Chrotopterus auritus* suggest that the diet of the studied group consists mainly of small vertebrates, accounting for nearly 80% of the total dry weight (including hair, feathers, and bones). This aligns with the expectations for a carnivorous bat (Gual-Suárez and Medellín 2021) and with prior reports on *C. auritus* as a predator that primarily consumes vertebrates of various sizes, supplementing its diet with insects year-round (de Oliveira et al. 2017).

Medellin (1989) described *C. auritus* as primarily carnivorous and insectivorous. Reported prey includes lizards, birds (e.g., *Columbina* spp.), small mammals like mouse opossums (*Marmosa* spp.), shrews (*Sorex* spp.), and mice



Table 2 Abundance (number of items in a given category), richness (number of taxa in the season) and relative frequency (percentage of a given taxon in the samples) of the taxa found in the samples and carcasses left by a group of *Chrotopterus auritus* over one year

Item Category	Family	Taxon*	Seasonal abundance of each taxon			
			Windy Season	Dry Season	Rainy Season	Abundance per taxon (relative frequency %)***
Mammals	Phyllostomidae	Desmodus rotundus ^{a,c,d}	1		8	9 (4.33)
		Artibeus jamaicensis ^{a,b,c,d}	_	_	1	1 (0.48)
	Mormoopidae	Mormoopidae sp. 1 ^{a,d}	_	_	1	1 (0.48)
	Soricidae	Soricidae spp. a,b,d	_	1	1	2 (0.96)
		Cryptotis spp. a,b,d	3	1	1	5 (2.40)
		Cryptotis nelson ^{a,b,d}	2	1	2	5 (2.40)
	Cricetidae	Rodentia sp. ^{a,b}	3	1	1	5 (2.40)
		Peromyscus mexicanus ^{a,b,c,d}	1	_	2	3 (1.44)
		Peromyscus leucopus ^{a,b,c,d}	_	_	1	1 (0.48)
		Peromyscus sp. a,b,c,d	_	_	1	1 (0.48)
		Nyctomys sp. 1 a,b,c,d	1	_	1	2 (0.96)
		Oryzomys alfaroi ^{a,b,c,d}	_	1	1	2 (0.96)
Birds	Columbidae	Columbidae sp. 1 ^{a,e}	2	1	9	12 (5.77)
		Columbina sp. a,e	4	1	6	11 (5.29)
	Thraupidae	Cyanerpes cyaneus ^e	_	_	6	6 (2.89)
	Trochilidae	Trochilidae sp. 1 ^e	1	1	2	4 (1.92)
Insects	Cerambycidae	Cerambycidae sp. 1 ^f	8	9	22	39 (18.75)
	Chrysomelidae	Chrysomelidae sp. 1 ^f	1	_	_	1 (0.48)
	Passalidae	Passalidae sp. 1 ^f	9	10	20	39 (18.75)
	Scarabaeidae	Scarabaeidae sp. 1 ^f	_	_	1	1 (0.48)
	Staphylinidae	Staphylinidae sp. 1 ^f	_	1	_	1 (0.48)
		Orthoptera sp 1 ^f	3	13	39	55 (26.44)
Abundance/season			39	41	126	208

^{*}Type of remains used for identification (as bold superindexes): a) bones (mandibles, radius, etc.), b) teeth, c) hair, d) whole body parts (head, limbs, tails), e) feathers, f) exoskeleton pieces (mainly legs) and arthropod wings

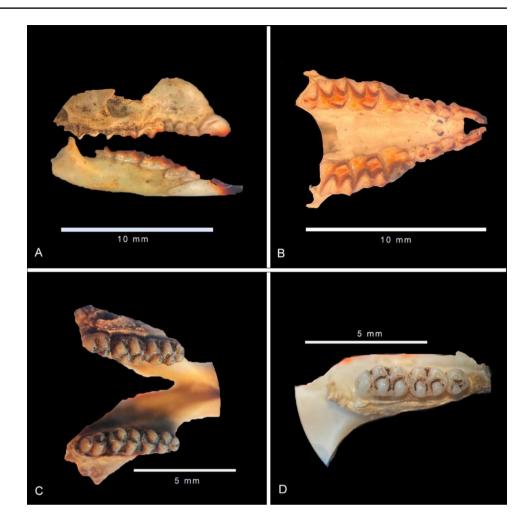
(Heteromys goldmani, Peromyscus oaxacensis, Nyctomys sumichrasti, and Ototylomys phyllotis). Insects consumed include families from the Coleoptera order. Vleut et al. (2019) noted a similar diet, however, this species also consumes some fruits (Uieda et al. 2007; Lintulaakso et al. 2023; Carballo-Morales et al. 2024). Our findings also identified mammals such as Peromyscus, Oryzomys, Nyctomys, and Desmodus as part of the diet, with insects as the main food category year-round, and birds, particularly chicks. No direct evidence of plant consumption was found. The absence of lizards or larger mammals may be due to differences in species availability between our study and those done in the south of the country (i.e., Calakmul in Campeche and El Triunfo in Chiapas, both with similar vegetation and climate to Los Tuxtlas, but less fragmented forests).

Due to the scattered, incomplete remains found at the roost, we classified most prey at the morphospecies level. The reported biomass reflects only identifiable remains, such as the discarded or rejected parts, including faces, skulls, beaks, bones, limbs, and long feathers. Viscera are also frequently discarded, as reported by Medellín (1989) in his study. Insect remains were mostly legs and wings, while mammal remains were typically fragmented jaws or palates with worn-out teeth, as stated by Alvarez et al. (2015). Feathers were abundant but mostly unidentifiable juvenile feathers. Among the mammal remains, rodent species such as Oryzomys and Peromyscus were prevalent, consistent with Witt and Fabian's (2010) observations in Brazil, where Rodentia made up 47.61% of the total sample. Cryptotis nelsoni, a shrew species endemic to Los Tuxtlas, was found in the diet, marking a new dietary record for C. auritus.



^{**}Percentage (in parenthesis, rounded to the upper unit)

Fig. 2 Cranium remains found among the items preyed upon by the studied *C. auritus* group over one year: A) Palate and jaw of *Cryptotis* sp.; B) Palate of *Cryptotis* sp.; C) Palate of *Peromyscus leucopus*; D) Mollar teeth of *Peromyscus mexicanus*



Previously documented prey species included Glossophaga soricina, Carollia perspicillata, Peropteryx macrotis, Tadarida brasiliensis, and Myotis sp. (Acosta 1951; Arita and Vargas 1995; Bonato et al. 2004; Bordignon 2005). In this study, we identified a mormoopid bat (likely Pteronotus mesoamericanus), Artibeus jamaicensis, and Desmodus rotundus, all of which share roost with this C. auritus group. This supports the idea of C. auritus as an opportunistic predator, potentially preying on roost-mates (at least in September).

Three bird groups were identified in the study: Passeriformes (*Cyanerpes cyaneus*), Apodiformes (Trochilidae), and Columbiformes (Columbidae). Medellin (1989) recorded a preference in *C. auritus* for Passeriformes, whereas *Vampyrum spectrum* favors larger bird species. In the studied family group, however, larger bird species were preferred. The most common bird remains were juvenile feathers, suggesting that chicks of various species were included in the diet of this *C. auritus* group, as large clumps of juvenile feathers accounted for 20.18% of the dry weight. In the Los Tuxtlas region, 17 species of columbids have been recorded (Monterrubio et al. 2016), 13 of which are

residents (Schaldach and Escalante-Pliego 1997). Feathers found across all three seasons likely belong to one of the 13 resident species in the region. In general, we found an increase in bird occurrence from March to August, mainly coinciding with the mating season of many birds in the study area (P.A. A.-R. pers. Obs.), suggesting that *C. auritus* may be taking advantage of the abundance of young birds. For example, *Cyanerpes cyaneus* breeding season varies depending on its geographical distribution (Peterson and Chalif 1989; Howell and Webb 1995; Herverth et al. 2016; Hilty 2018). In this study, feathers from this species were observed only during the rainy season (June to October), coinciding with the breeding season of the species reported for Mexico.

Of the 17 Trochilidae species in Los Tuxtlas (Wetmore 1943; Andrle 1967), Toledo (1975) identified 11 as permanent or occasional inhabitants of the study area, with four typically residing within the forest interior. Given that *C. auritus* is linked to conserved areas (Vleut et al. 2019) and may avoid disturbed regions (but still tolerant to habitat perturbation; see Gamboa Alurralde and Díaz 2021), the hummingbirds in its diet are likely *Phaethornis*



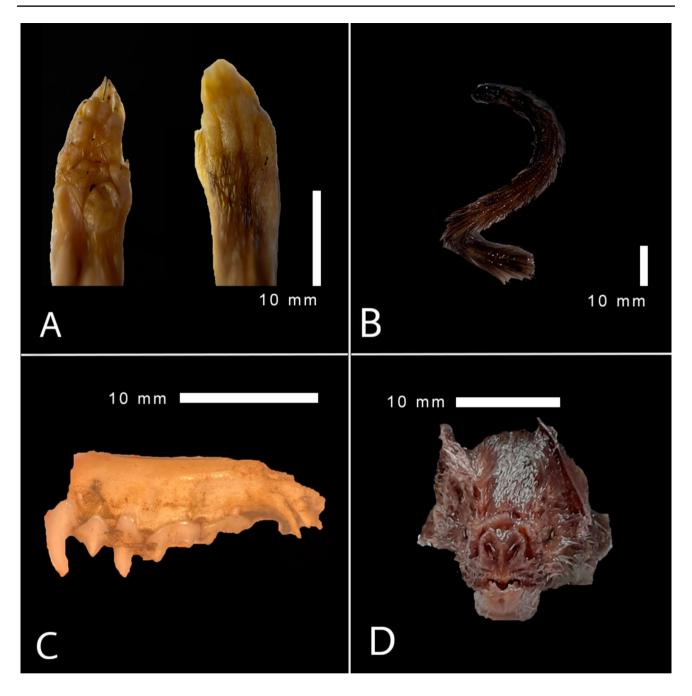


Fig. 3 Mammalian remains found among the prey of a *C. auritus* group: **A)** Dorsal (right) and ventral (left) views of a foot of *Oryzomys alfaroi*; **B)** Severed tail of *Nyctomys* sp.; **C)** Superior teeth of *Artibeus jamaicensis*; **D)** Frontal view of a severed head of *Desmodus rotundus*

superciliosus, Campylopterus, Amazilia tzacat, or Campylopterus curvipennis.

Comparing the diet of *C. auritus* with its relative *Vampyrum spectrum*, prey sizes for the latter range from 20 to 150 g (Vehrencamp et al. 1977), overlapping with the *C. auritus* range of 10 to 35 g, and up to 70 g (Medellín 1988). *V. spectrum* typically captures larger avian prey and some bats (McCarthy 1987), while *C. auritus* primarily consumes rodents and small birds, rarely preying on bats (Medellin

1989). This study reflects this pattern, with bat remains found only in September.

Regarding seasonal variation, we observed the highest abundance of food items during rainy season (Table 2). Bats' diets may reflect prey availability (Burles et al. 2008), especially during rainy season (Bonaccorso 1979; Nurul-Ain et al. 2017). Additionally, previous studies show bats change their diet during pregnancy and lactation (Kunz 1974; Anthony and Kunz 1977; McLean and Speakman



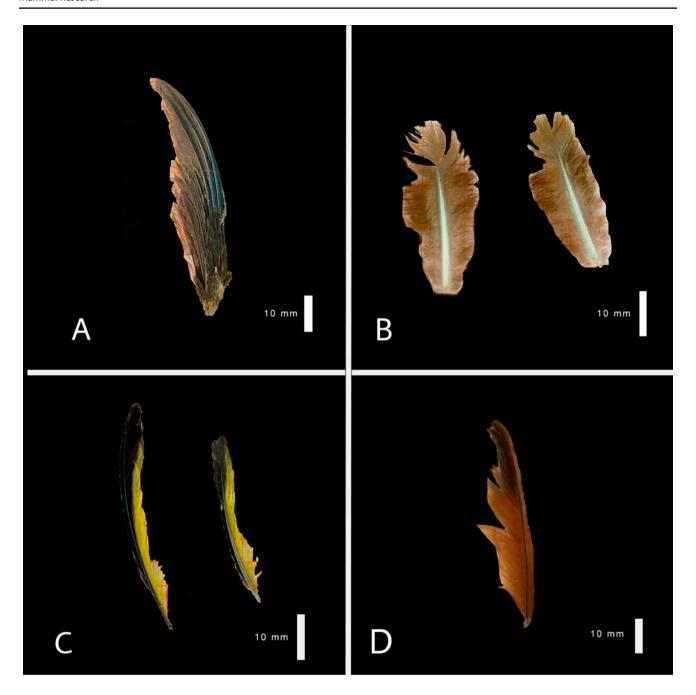


Fig. 4 Some feathers found among the food remains in the studied *C. auritus* group: **A**) Feather of *Columbina* sp.; **B**) Feathers of Columbidae; **C**) Feathers of *Cyanerpes cyaneus*; **D**) Feather of Trochilidae

1999; Haarsma et al. 2023). Lactating females also require more calcium (Kovacs and Kronenberg 1997), and insects are poor sources of this mineral (Booher and Hood 2010).

Mammal intake remained consistent year-round in this study, while bird consumption rose during dry and rainy seasons, corresponding with pregnancy and lactation, and dropped in the windy season; this could also be due to the reproductive seasons of these bird species (Fig. 1). Insect remains were more abundant in the rainy season (higher dry

weight) and more diverse in the windy season. Insectivorous bats consume more insects during the breeding season due to high energy demands (Arango-Diago et al. 2020), and this reasoning can be applied to *C. auritus's* breeding season. The species has a gestation period of approximately seven months (Taddei 1976), with mating estimated between August and September (the rainy season at the study site) (Álvarez et al. 2015, 2018) and births in April and May (dry season at the site) (Wilson 1979; Álvarez-Yax et al. 2018).



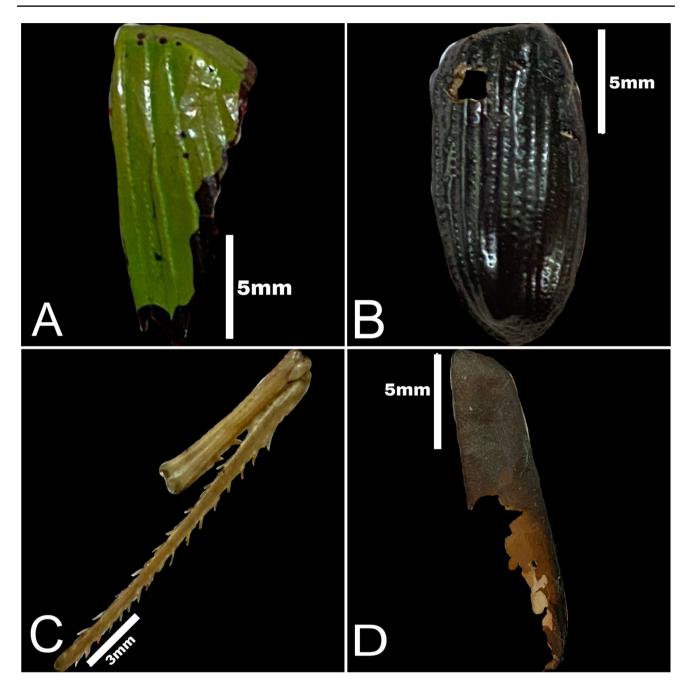


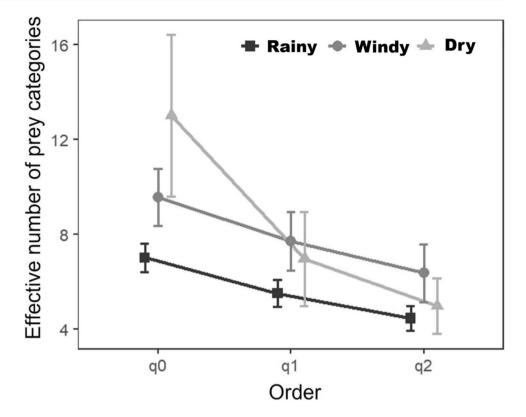
Fig. 5 Insect remains found among the items consumed by the studied *C. auritus* group: **A**) Elytra of Scarabaeidae; **B**) Wing of Passalidae; **C**) Leg of Orthoptera; **D**) Elytra of Cerambycidae

Results show an increase in dry-weight biomass during the rainy season (83.37% of the total sample), though this coincides with the lowest 0D diversity by category (i.e., fewer morphospecies). Additionally, the rainy season had the highest mammalian remains and coincided with observations of a lactating female in the group (May to July; Fig. 1), possibly reflecting increased intake to support the energy needs of lactation (Kunz et al. 1995). Prey items variability may also be related to the age and handling experience of

juveniles. By our sampling design, we were unable to distinguish between preys consumed by adults and by juveniles, but Phyllostomid bats, including carnivorous species, learn prey selection and handling from their mothers (Vehrencamp et al. 1977; Patriquin and Ratcliffe 2023), and juveniles tend to consume smaller and easier-to-capture prey (Aldasoro et al. 2024). Bigger prey remains (like full heads and forearms from *Desmodus* and *Artibeus* bats) were present in the samples after the rearing of the small pup.



Fig. 6 Diversity of prey from *Chrotopterus auritus* in the three seasons for effective species in the orders: q0 (richness), q1 (abundant species) and q2 (very abundant species). Diversity of prey items peaks in the dry season, in all three components



This study, which examined a single-family group of up to four C. auritus individuals (a mating pair and two juveniles; Video S1), provides the first year-long analysis of dietary remains for this species. It also represents the northernmost diet report for C. auritus, documenting new food items not previously recorded in other areas, including Cryptotis nelsoni, endemic to the area. Furthermore, species such as *Peromyscus mexicanus*, *Peromyscus leucopus*, Oryzomys alfaroi, and Cyanerpes cyaneus were added to the known diet of C. auritus. Few studies address carnivorous bat diets across geographic areas, though some Phyllostomidae show dietary shifts across regions (e.g., species within Trachops cirrhosus complex and Artibeus jamaicensis; Tuttle and Ryan 1981; Kalko et al. 1996; Bonato and Facure 2000; Estrada et al. 1984; Ortega and Castro-Arellano 2001; Genoways et al. 2005; Lobova et al. 2009). Geographic variation in prey availability may also apply to C. auritus, as evidenced by prey differences within its distribution range, such as the predation of *C. nelsoni*, exclusively in Los Tuxtlas.

Our results confirm that *C. auritus* is a carnivorous bat in this region, relying on small vertebrates, mainly birds and mammals, for over 70% of its diet. Additionally, the diet of *C. auritus* varies throughout the year, likely reflecting the reproductive cycles and availability of its prey. These changes lead to increased consumption of certain organisms due to high energy demands and fluctuations in resource availability, aligning with previous studies on carnivorous bats and *C. auritus*. Unlike the southern populations of *C.*

auritus in Mexico, where multiple roosts are known (Vleut et al. 2019; Gual-Suárez et al. 2025), this is the only known roost in Los Tuxtlas. Additional studies in other roosts and months, ideally incorporating DNA analysis from guano and prey remains, would help determine whether these dietary trends are consistent across the species range.

It was expected that the rainy season would show the highest dietary diversity compared to the dry and windy seasons. However, this was not confirmed, as there was no significant difference in dietary diversity between the three seasons. Still, the rainy season did record the highest biomass in grams, indicating that during this season, *C. auritus* consumes a larger quantity of food but from a more limited range of species, suggesting a selective diet. For component q0, the most prevalent species appeared in the dry season; for q1 and q2, the most prevalent species were recorded during the windy and dry seasons. Overall, the rainy season had the lowest prevalence of certain species, suggesting that *C. auritus* consumes more common species in the rainy season, while in the dry and windy seasons, it may opportunistically consume rarer species when available.

Trophic guild and niche breadth reflect habitat heterogeneity and are related to extinction risk in Phyllostomidae (Carballo-Morales et al. 2024). Therefore, understanding diet is crucial for conservation, particularly in heavily deforested regions like Los Tuxtlas (Dirzo and García 1992; Von Thaden et al. 2018). This study also reflects the species plasticity and its potential to persist in small patches



of well-preserved forest matrix, provided suitable roosts (Vleut et al. 2019; Gual-Suárez et al. 2025) and prey are available (but see Shaw et al. 2013). The presence of *C. auritus*, a top-level predator, suggests the continued presence of its prey and indicates a functioning ecosystem.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s13364-025-00804-2.

Acknowledgements We sincerely thank Benigno Absalon for his help during fieldwork and Jose Alberto Lobato García for his assistance in identifying bird species. SMCJ thanks Dirección General de Investigaciones at Universidad Veracruzana for the SNI fellowship.

Author contributions The study was conceived by PAAR. Fieldwork and sample collection were carried out by PAAR, MAM and MCMG. All sample preparation and initial data analysis were done by SMCJ and PAAR. The first draft of the manuscript was written by SMCJ and PAAR. All authors contributed to the data analysis, as well as manuscript correction. All authors read and approved the final version of the manuscript.

Funding This research received no external funding.

Data availability The data that supports the findings of this study is available from the corresponding author upon reasonable request.

Declarations

Competing interests The authors declare that they have no conflict of interest.

References

- Acosta-Lara EF (1951) Notas ecológicas sobre algunos quirópteros del Brasil. Comunicaciones Zoológicas Del Museo De Montevideo 3(65):1–2
- Aldasoro M, Vallejo N, Olasagasti L, Diaz de Cerio O, Aihartza J (2024) Learning to hunt on the go: dietary changes during development of Rhinolophid bats. Animals 14(22):3303. https://doi.org/10.3390/ani14223303
- Altenbach JS (1979) Locomotor Morphology of the Vampire Bat, *Desmodus rotundus*. Special Publication no. 6. The American Society of Mammalogists, Pittsburgh, PA. https://doi.org/10.5962/bhl.title.39538
- Álvarez ST, Álvarez T, González N (2015) Guía para la identificación de los mamíferos de México en campo y laboratorio. Asociación Mexicana de Mastozoología Guadalajara Mexico:522
- Álvarez-Yax RA, Gómez-Lemus AH, Hernández-Fuentes JS, Juárez-Bolaños AP, Pérez-Quan KJ, Tijerino-Escobar DG, Villatoro-Castañeda M, Ariano-Sánchez D (2018) Reproducción del falso vampiro lanudo *Chrotopterus auritus* (Chiroptera: Phyllostomidae) en un bosque tropical húmedo de la costa pacífica de Guatemala. Acta Zool Mex 34:1–3. https://doi.org/10.21829/azm. 2018.3411190
- Andrle FR (1967) Birds of the Sierra de Tuxtla in Veracruz, Mexico. Wilson Bull 79(2):163–187
- Anthony EL, Kunz TH (1977) Feeding strategies of the little brown bat, *Myotis lucifugus*, in Southern New Hampshire. Ecology 13:775–786
- Arango-Diago S, Castillo-Figueroa D, Albarracín-Caro J, Pérez-Torres J (2020) Dietary variation and reproductive status of

- Mormoops megalophylla (Chiroptera: Mormoopidae) in a cave of northeastern Andes from Colombia. Mastozoología Neotropical 27(2):258–265. https://doi.org/10.31687/saremMN. 20.27.2.0.13
- Araujo D, Machado M (2012) Ampliación del límite altitudinal de *Chrotopterus auritus* Peter, 1856 (Mammalia: Chiroptera) en Venezuela y algunos comentarios ecológicos. Ecotrópicos 25(1):35–38. https://doi.org/10.53157/88tj-gwpd
- Arita HT, Vargas JA (1995) Natural history, interspecific association, and incidence of the cave bats of Yucatan, Mexico. Southw Naturalist 40(1):29–37
- Arnett R Jr, Thomas MC, Skelley PE, Frank JH (2002) American Beetles. Scarabaeoidea through Curculionoidea. CRC Press. Boca Raton, Fl, Polyphaga, p 860
- Avibase. Mielerito patirrojo *Cyanerpes cyaneus* (Linnaeus, 1766)». Avibase. https://avibase.bsc-eoc.org
- Baptista F, Trail P, Horblit H (1997) Familia Columbidae (palomas y palomos). In: Del Hoyo J, Elliott A, Sargatal J (eds) Manual de aves del mundo. Lynx Edicions, Barcelona
- Barquez RM, Perez S, Miller B, Díaz MM (2015) Chrotopterus auritus. In: The IUCN Red List of Threatened Species, vol 2015, p e.T4811A22042605. https://doi.org/10.2305/IUCN.UK.2015-4. RLTS.T4811A22042605.en
- Bernis F, De Juana E, Del Hoyo J, Fernández-Cruz M, Ferrer X, Sáez-Royuela R, Sargatal J (1998) Nombres en castellano de las aves del mundo recomendados por la Sociedad Española de Ornitología (Pterocliformes, Columbiformes, Psittaciformes y Cuculiformes). Ardeola 45(1):87–96
- Bonaccorso F (1979) Foraging and reproductive ecology in a Panamanian bat community. Bull Fla Mus. Nat Hist 24(4):359–408. https://doi.org/10.58782/flmnh.dobh1085
- Bonato V, Facure K (2000) Bat predation by the fringe-lipped bat *Trachops cirrhosus* (Chiroptera: Phyllostomidae). Mammalia 64:241–243
- Bonato V, Gomes K, Uieda W (2004) Food habits of bats of subfamily Vampyrinae in Brazil. J Mammal 85:708–713. https://doi.org/10.1644/BWG-121
- Booher CM, Hood WR (2010) Calcium utilization during reproduction in big brown bats (*Eptesicus fuscus*). J Mammal 91(4):952–959. https://doi.org/10.1644/09-MAMM-A-186.1
- Bordignon MO (2005) Predação de morcegos por *Chrotopterus auritus* (Peters) (Mammalia, Chiroptera) no pantanal de Mato Grosso do Sul. Brasil Rev Bras Zool 22(4):1207–1208. https://doi.org/10. 1590/s0101-81752005000400058
- Borloti I, Pimenta T, Ditchfield A (2019) First record of predation of Nyctinomops laticaudatus (É. Geoffroy, 1805) by Chrotopterus auritus (Peters, 1856) (Mammalia: Chiroptera). Biodivers Data J:e38303. https://doi.org/10.3897/BDJ.7.e33829
- Boyles JG, Storm JJ (2007) The perils of picky eating: dietary breadth is related to extinction risk in insectivorous bats. PLoS ONE 2(7):e672. https://doi.org/10.1371/journal.pone.0000672
- Brito JEC, Miranda JMD, Bernardi IP, Passos FC (2010) Predação de *Tadarida brasiliensis* por *Chrotopterus auritus* no sul do Brasil. Chiroptera Neotrop 16(Suppl. 1):45–46
- Burles DW, Brigham RM, Ring RA, Reimchen TE (2008) Diet of two insectivorous bats, *Myotis lucifugus* and *Myotis keenii*, in relation to arthropod abundance in a temperate Pacific Northwest rainforest environment. Can J Zool 86:1367–1375. https://doi.org/10.1139/Z08-125
- Carballo-Morales JD, Villalobos F, Saldaña-Vázquez RA, Herrera-Alsina L (2024) The habitat breadth of phyllostomid bats is partially determined by their diet and could be used as a predictor of extinction risk. Biodivers Conserv 33:3129–3144. https://doi.org/10.1007/s10531-024-02905-x
- Carvalho F, Bôlla DAS, Mottin V, Kiem SZ, Zocche JJ, Passos FC (2020) Chilling to the bone: Lower temperatures increase



- vertebrate predation by *Tonatia bidens* (Chiroptera: Phyllostomidae). Zoologia 37:1–5. https://doi.org/10.3897/zoologia.37. e37682
- Castillo-Campos G, Laborde J (2004) La vegetación. In: Guevara S, Laborde DJ, Sánchez-Ríos G (eds) Los Tuxtlas. El paisaje de la sierra. Instituto de Ecología and Unión Europea, pp 231–265
- Ceballos G, Oliva L (2002) Los mamíferos silvestres de México. Fondo de Cultura Económica, España, p 986
- Chao A, Gotelli NJ, Hsieh TC, Sander EL, Colwell RK, Ma KH, Ellison AM (2014) Rarefaction and extrapolation with Hill numbers: a framework for sampling and estimation in species diversity studies. Ecol Monogr 84(1):45–67. https://doi.org/10.1890/13-0133.1
- Choate JR (1973) Cryptotis mexicana. Mamm Spec 28:1–2. https://doi.org/10.2307/0.28.1
- Cory CB (1918) A Catalogue of birds of the Americas. Field Museum Natural History, Zoological Series 13(197):1–315
- Da Silva FB, Soto-Centeno JA, Simmons NB, Ditchfield AD, Leite YLR (2024) A species complex in the iconic frog-eating bat *Trachops cirrhosus* (Chiroptera, Phyllostomidae) with high variation in the heart of the Neotropics. Am Mus Novit 4021(1):1–17. https://doi.org/10.1206/4021.1
- de Oliveira HFM, de Camargo NF, Gager Y, Aguiar LMS (2017) The response of bats (Mammalia: Chiroptera) to habitat modification in a Neotropical savannah. Trop Conserv Sci 10. https://doi.org/10.1177/1940082917697263
- Díaz MM, Solari S, Gregorín R, Aguirre LF, Barquez R (2021) Clave de identificación de los murciélagos Neotropicales/Chave de indentifição dos morcegos Neotropicais, vol 4. Publicación Especial PCMA (Programa de Conservación de los Murciélagos de Argentina), Bilingüe: español-portugués, p 211
- Dirzo R, Garcia MC (1992) Rates of deforestation in Los Tuxtlas, a Neotropical area in Southeast Mexico. Conserv Biol 6(1):84–90. https://doi.org/10.1046/j.1523-1739.1992.610084.x
- Estrada A, Coates-Estrada R, Vasquez-Yanes C, Orozco-Segovia A (1984) Comparison of frugivory by howling monkeys (*Alouatta palliata*) and bats (*Artibeus jamaicensis*) in the tropical rain forest of Los Tuxtlas, Mexico. Am J Primatol 7(1):3–13. https://doi.org/10.1002/ajp.1350070103
- Felix S, Morim Novaes RL, de França SR, Tadeu Santori R (2013) Diet of *Tonatia bidens* (Chiroptera, Phyllostomidae) in an Atlantic Forest area, southeastern Brazil: first evidence for frugivory. Mammalia 77(451):454. https://doi.org/10.1515/mammalia-2012-0117feliz
- Gamboa Alurralde S, Díaz MM (2021) Assemblage-level responses of Neotropical bats to forest loss and fragmentation. BAAE 50:57– 66. https://doi.org/10.1016/j.baae.2020.09.001
- Gardner A (2008) Mammals of South America, Marsupials, Xenarthrans, Shrews, and Bats. Volume 1. The University of Chicago Press, Chicago, Illinois, p 690
- Gardner A (1977) Feeding habits. In: Biology of bats of the new world family phyllostomidae. Special Publications, The Museum of Texas Tech University, Texas, pp 1–364. https://doi.org/10.5962/bhl.title.142603
- Genoways HH, Baker RJ, Bickham JW, Phillips CJ (2005) Bats of Jamaica. Spec Publ Mus Texas Tech Univ 48:1–154. https://doi.org/10.5962/bhl.title.142604
- Gual-Suárez F, Medellín RA (2021) We eat meat: a review of carnivory in bats. Mamm Rev 51(4):540–558. https://doi.org/10.1111/mam.12254
- Gual-Suárez F, Trujillo LA, Torres-Cervantes J, Ordóñez-García MC, Medellín RA (2025) Guardians of the Mayan temples: microclimatic conditions of archaeological sites used as roosts by *Chrotopterus auritus* in Southeastern Mexico. Mammal Notes 11(1):1–12. https://doi.org/10.47603/mano.v11n1.487

- Haarsma AJ, Jongejans E, Duijm E, van der Graaf C, Lammers Y, Sharma M, Siepel H, Gravendeel B (2023) Female pond bats hunt in other areas than males and consume lighter prey when pregnant. J Mammal 104(6):1191–1204. https://doi.org/10.1093/jmammal/gyad096
- Hall ER, Kelson KR (1959) The mammals of North America. The Ronald Press Co., New York, NY, p 1083
- Hansson L (1970) Methods of morphological diet microanalysis in rodents. Oikos 21(2):255–266. https://doi.org/10.2307/35436 82
- Herverth V, Padilla J, Garrido R, Martínez I (2016) Registros del Mielero pata roja (*Cyanerpes cyaneus*) en el estado de Hidalgo, México. Tecnointelecto 13:1–5
- Hill JE, Smith JD (1984) Bats, a natural history. In: British museum of natural history. University of Texas Press, Austin, TX, p 254
- Hilty S (2018) Red-legged Honeycreeper (*Cyanerpes cyaneus*). In: Handbook of the birds of the world alive. Lynx Edicions, Barcelona. https://doi.org/10.2173/bow.relhon1.01
- Howell SNG (2002) Hummingbirds of North America: The Photographic guide. Princeton University Press, San Diego, CA, p 272
- Howell SNG, Webb S (1995) A guide to the birds of Mexico and Northern Central America. Oxford University Press, New York, NY, p 851
- Hsieh TC, Ma KH, Chao A (2020) iNEXT: an R package for rarefaction and extrapolation of species diversity (Hill numbers). Methods Ecol Evol 7(12):1451–1456. https://doi.org/10.1111/2041-210X.12613
- Huckaby DG (1980) Species limits in the *Peromyscus mexicanus* group (Mammalia: Rodentia: Muroidea). Contrib Sci 326:1–24. https://doi.org/10.5962/p.241261
- Johnsgard PA (1997) The hummingbirds of North America, 2nd edn. Smithsonian Institution Scholarly Press, London, p 314
- Jordan MJR (2005) Dietary analysis for mammals and birds: a review of field techniques and animal-management applications. IntZoo Yearb 39(1):108–116. https://doi.org/10.1111/j.1748-1090.2005. tb00010.x
- Kalko E, Handley C, Handley D (1996) Organization, diversity, and long-term dynamics of a Neotropical bat community. In: Cody ML, Smallwood JA (eds) Long-term studies of vertebrate communities. Academic Press, Boston, MA, pp 503–553
- Kovacs CS, Kronenberg HM (1997) Maternal–fetal calcium and bone metabolism during pregnancy, puerperium, and lactation. Endocr Rev 18(6):832–872. https://doi.org/10.1210/edrv.18.6.0328
- Kunz TH (1974) Feeding ecology of a temperate insectivorous bat (*Myotis velifer*). Ecology 13:693–711
- Kunz TH, Whitaker JO Jr, Wadanoli MD (1995) Dietary energetics of the insectivorous Mexican free-tailed bat (Tadarida brasiliensis) during pregnancy and lactation. Oecologia 101(4):407–415. https://doi.org/10.1007/BF00317311
- Leschen R, Beutel RG, Lawrence J (2010) Handbook of Zoology. In: Morphology and Systematics (Ealetroidea, Bostrichiformia, Cucujjiformia partim), vol 2. Walter de Gruyter GmbH & Co., New York, NY, p 799
- Linares JO (1998) Mamíferos de Venezuela. Sociedad Conservacionista Audubon de Venezuela, Caracas, p 691
- Lintulaakso K, Tatti N, Žliobaitė I (2023) Quantifying mammalian diets. Mamm Biol 103:53–67. https://doi.org/10.1007/s42991-022-00323-6
- Lobova T, Geiselman CK, Mori SA (2009) Seed dispersal by bats in the Neotropics. The New York Botanical Garden, New York NY, p 465
- MacGregor-Fors I, Payton ME (2013) Contrasting diversity values: Statistical inferences based on overlapping confidence intervals. PLoS ONE 8(2):e56794. https://doi.org/10.1371/journal.pone. 0056794



- Martínez-Fonseca JG, Mau R, Walker FM, Medina-Fitoria A, Yasuda K, Chambers CL (2022) Vampyrum spectrum (Phyllostomidae) movement and prey revealed by radio-telemetry and DNA metabarcoding. PLoS ONE 17(4):e0265968. https://doi.org/10.1371/journal.pone.0265968
- McAney C, Shiel C, Sullivan C, Fairley J (1991) The analysis of bat droppings. Occas Publ Mamm Soc London 14(48):1–48
- McCarthy T (1987) Additional mammalian prey of the carnivorous bats, *Chrotopterus auritus* and *Vampyrum spectrum*. Bat Res News 28(1–2):1–3
- McLean JA, Speakman JR (1999) Energy budgets of lactating and non-reproductive Brown Long-Eared Bats (*Plecotus auritus*) suggest females use compensation in lactation. Funct Ecol 13(3):360–372. https://doi.org/10.1046/j.1365-2435.1999.00321.x
- Medellín RA (1988) Prey of *Chrotopterus auritus*, with notes on feeding behavior. J Mammal 69(4):841–844. https://doi.org/10.2307/1381644
- Medellín RA (1989) *Chrotopterus auritus*. Mamm Species 343:1–5. https://doi.org/10.2307/3504232
- Monterrubio-Rico TC, Villaseñor F, Álvarez-Jara M, Escalante P (2016) Ecología y situación actual de la familia Columbidae en la Reserva de la Biosfera de Los Tuxtlas, Veracruz. Ornitol Trop 27:17–26. https://doi.org/10.58843/ornneo.v27i0.124
- Navarro LD, Wilson DE (1982) Vampyrum spectrum. Mamm Species 184:1–4. https://doi.org/10.2307/3503798
- Nogueira M, Monteiro L, Peracchi A (2007) New evidence of bat predation by the Woolly False Vampire Bat *Chrotopterus auritus*. Chiroptera Neotrop 12(2):286–288
- Nurul-Ain E, Rosli H, Kingston T (2017) Resource availability and roosting ecology shape reproductive phenology of rain forest insectivorous bats. Biotropica 49(3):382–394. https://doi.org/10.1111/btp.12430
- Ochoa JO, Bevilacqua M, García F (2005) Evaluación ecológica rápida de las comunidades de mamíferos en cinco localidades del Delta del Orinoco, Venezuela. Interciencia 30:466–475
- Ortega J, Castro-Arrellano I (2001) Artibeus Jamaicensis. Mamm Species 662:1–9. https://doi.org/10.2307/3504520
- Patriquin KJ, Ratcliffe JM (2023) Bats learn about potential food sources from others: a review. Can J Zool 101:294–306. https://doi.org/10.1139/cjz-2022-0119
- Peterson RT, Chalif EL (1989) Aves de México. Editorial Diana, Mexico City, p 473
- Reid FA (2009) A field guide to the mammals of Central America and Southeast Mexico, 2nd edn. Oxford University Press, New York, p 368
- Santos M, Aguirre LF, Vázquez LB, Ortega J (2003) Phyllostomus hastatus. Mamm Species 722:1–6. https://doi.org/10.1644/0.722.1
- Schaldach WJ Jr, Escalante-Pliego P (1997) Lista de aves. In: González Soriano E, Dirzo R, Vogt RC (eds) Historia natural de Los Tuxtlas. Instituto de Biología and Instituto de Ecología, Universidad Autónoma de México/ CONABIO, México D.F. Mexico, pp 571–588
- Schodde R, Mason IJ (1997) Aves (Columbidae a Coraciidae). CSIRO Publishing, Clayton
- SEMARNAT (Secretaría de Medio Ambiente y Recursos Naturales) (2010) Norma Oficial Mexicana NOM-059-SEMAR-NAT-2010, Protección ambiental especies nativas de México de flora y fauna silvestres. Diario Oficial de la Federación del 30 de diciembre de 2010, Distrito Federal, Mexico.
- Shaw DW, Escalante P, Rappole JH, Ramos MA, Oehlenschlager RJ, Warner DW, Winker K (2013) Decadal changes and delayed avian species losses due to deforestation in the northern Neotropics. PeerJ:e179. https://doi.org/10.7717/peerj.179
- Simmons N, Cirranello A (2025) Bat species of the world: a taxonomic and geographic database. https://www.mammalwatching.com/2020/03/25/bats-of-the-world-database/

- Slud P (1964) The birds of Costa Rica: distribution and ecology. Bull Am Mus Nat Hist 128:5–430
- Soto M (2004) Clima. In: Guevara S, Laborde J, Sánchez-Ríos G (eds) Los Tuxtlas: el paisaje de la sierra. Instituto de Ecología A. C, Xalapa, pp 195–199
- Taddei VA (1976) The reproduction of some Phyllostomidae (Chiroptera) from the northwestern region of the state of Sao Paulo. Bol Zool Univ Sao Paulo 1:313–330. https://doi.org/10.11606/issn. 2526-3358.bolzoo.1976.121587
- Toledo VM (1975) La estacionalidad de las flores utilizadas por los colibríes de una selva tropical húmeda en México. Biotropica 7(1):63–70. https://doi.org/10.2307/2989802
- Tuttle MD, Ryan MJ (1981) Bat predation and the evolution of frog vocalization in the Neotropics. Science 214(4521):677–678. https://doi.org/10.1126/science.214.4521.677
- Uieda W, Sato TM, De Carvalho MC, Bonato V (2007) Fruits as unusual food items of the carnivorous bat *Chrotopterus auritus* (Mammalia: Phyllostomidae) from southeastern Brazil. Rev Bras Zool 24(3):844–847. https://doi.org/10.1590/S0101-8175200700 0300035
- Vehrencamp SL, Stiles FG, Bradbury JW (1977) Observations on the foraging behavior and avian prey of the neotropical carnivorous bat. Vampyrum Spectrum J Mamm Evol 58(4):469–478. https:// doi.org/10.2307/1379995
- Vleut I, Carter GG, Medellín RA (2019) Movement ecology of the carnivorous woolly false vampire bat (*Chrotopterus auritus*) in southern Mexico. PLoS ONE 14:e0220152. https://doi.org/10. 1371/journal.pone.0220152
- Von Thaden JJ, Laborde J, Guevara S, Venegas-Barrera CS (2018) Forest cover change in the Los Tuxtlas biosphere reserve and its future: the contribution of the 1998 protected natural area decree. Land Use Pol 72:443–450. https://doi.org/10.1016/j.landusepol. 2017.12.040
- Wetmore A (1943) The birds of southern Veracruz, Mexico. Proc US Natl Mus 93(3164):215–340. https://doi.org/10.5479/si.00963 801.93-3164.215
- Whitaker JO Jr, Maser C, Cross SP (1981) Food habits of eastern Oregon bats, based on stomach and scat analysis. Northwest Sci 55(4):281–292
- Whitaker JO Jr, McCracken G, Siemers B (2009) Food habits analysis of insectivorous bats. In: Kunz TH, Parsons S (eds) Ecological and behavioral methods for the study of bats. Johns Hopkins University Press, Baltimore, MD, pp 567–592
- White RE (1983) A field guide to the beetles of North America. Houghton Mifflin Harcourt, Boston, MA, p 384
- Wilson D (1979) Reproductive patterns. In: Baker RJ, Jones JK, Carter DC (eds) Biology of bats of the New World family Phyllostomidae. Part III. Special Publications of the Museum of Texas Tech University, Lubbock, TX, pp 317–378
- Witt AA, Fabián ME (2010) Hábitos alimentares e uso de abrigos por Chrotopterus auritus (Chiroptera, Phyllostomidae). Mastozoología Neotrop 17(2):353–360
- Woodman N (2018) American recent Eulipotyphla: Nesophontids solenodons moles and shrews in the New World. USGS Publications. Smithsonian Institution Scholarly Press, Washington D. C, p 107
- **Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.
- Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

