

First record of bat-pollination in the species-rich genus *Tillandsia* (Bromeliaceae)

Pedro Adrián Aguilar-Rodríguez^{1,*}, M. Cristina MacSwiney G.¹, Thorsten Krömer¹,
José G. García-Franco², Anina Knauer³ and Michael Kessler³

¹Centro de Investigaciones Tropicales, Universidad Veracruzana, Casco de la ExHacienda Lucas Martín, Privada de Araucarias S/N. Col. Periodistas, C.P. 91019, Xalapa, Veracruz, México, ²Red de Ecología Funcional, Instituto de Ecología, A.C., Carretera antigua a Coatepec No. 351, El Haya, C.P. 91070, Xalapa, Veracruz, México and ³Institute of Systematic Botany, University of Zurich, Zollikerstrasse 107, CH-8008 Zurich, Switzerland

* For correspondence. E-mail pedroaguilarr@gmail.com

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- **Background and Aims** Bromeliaceae is a species-rich neotropical plant family that uses a variety of pollinators, principally vertebrates. *Tillandsia* is the most diverse genus, and includes more than one-third of all bromeliad species. Within this genus, the majority of species rely on diurnal pollination by hummingbirds; however, the flowers of some *Tillandsia* species show some characteristics typical for pollination by nocturnal animals, particularly bats and moths. In this study an examination is made of the floral and reproductive biology of the epiphytic bromeliad *Tillandsia macropetala* in a fragment of humid montane forest in central Veracruz, Mexico.
- **Methods** The reproductive system of the species, duration of anthesis, production of nectar and floral scent, as well as diurnal and nocturnal floral visitors and their effectiveness in pollination were determined.
- **Key Results** *Tillandsia macropetala* is a self-compatible species that achieves a higher fruit production through outcrossing. Nectar production is restricted to the night, and only nocturnal visits result in the development of fruits. The most frequent visitor (75 % of visits) and the only pollinator of this bromeliad (in 96 % of visits) was the nectarivorous bat *Anoura geoffroyi* (Phyllostomidae: Glossophaginae).
- **Conclusions** This is the first report of chiropterophily within the genus *Tillandsia*. The results on the pollination biology of this bromeliad suggest an ongoing evolutionary switch from pollination by birds or moths to bats.

Key words: *Tillandsia macropetala*, Bromeliaceae, bat-pollination, *Anoura geoffroyi*, bromeliad, chiropterophily, humid montane forest, floral visitors, Mexico, pollinator effectiveness.

INTRODUCTION

The family Bromeliaceae is distributed almost exclusively in the Neotropics (Benzing, 2000), and 56 % of its species are epiphytic (Zotz, 2013). The family comprises about 3160 species in 50 genera, and *Tillandsia* is the most diverse genus, with more than 670 species, of which 635 are epiphytic (Zotz, 2013). Among bromeliads, pollination by vertebrates predominates over that provided by insects, and most species are pollinated by hummingbirds (Kessler and Krömer, 2000; Canela and Sazima, 2005; Krömer *et al.*, 2006). In some bromeliad genera (e.g. *Guzmania*, *Pitcairnia* and *Vriesea*), however, syndromes of pollination by insects, birds and bats have evolved independently (Benzing, 2000). In particular, species of the subfamily Tillandsioideae (e.g. *Tillandsia*, *Guzmania* and *Vriesea*) have on various occasions modified their floral characteristics to attract a wide range of pollinators (Benzing, 2000).

The epiphytic bromeliad *Tillandsia macropetala* Wawra, together with *T. grandis* Schltdl. and *T. viridiflora* (Beer) Baker, belongs to the Mexican species in the *Tillandsia viridiflora* complex, among which *T. macropetala* was recently recognized as a species distinct from *T. viridiflora* (Krömer *et al.*, 2012). It is distributed in central and southern Mexico, in wet montane or pine–oak forests at elevations of 1100–2500 m. In contrast to the majority of species of the genus, *T. macropetala* does not present the conspicuously coloured bracts with

contrasting flowers and diurnal anthesis typical of pollination by hummingbirds (Endress, 1994; Wilmer, 2011). Instead, this species has a light-green corolla, green bracts and crepuscular anthesis (Krömer *et al.*, 2012).

Tillandsia macropetala was not included (even under its previous name *T. viridiflora*) in the classic work of Gardner (1986) about the pollination in *Tillandsia*, but the related species *T. heterophylla* E. Morren was catalogued as being moth-pollinated. These three species present particular cases within the genus *Tillandsia*, given their nocturnal anthesis and light-green petals, as seen in other bromeliads of the subgenus *Pseudoalcantarea* (Benzing, 2000). Whereas Hietz and Hietz-Seifert (1994) suggested that sphingid moths could be pollinators of *T. viridiflora*, Benzing (2000) argued based on their floral characteristics that the species of *Pseudoalcantarea* could be pollinated by bats.

The discovery of dilute nectar, rich in hexoses, in *T. viridiflora* (Krömer *et al.*, 2008), together with other observations also including *T. macropetala* (T. Krömer and M. C. MacSwiney G., pers. obs.), suggests that bats could be their principal pollinators. In addition, the helicoiform flowers of *T. macropetala* stay open during the morning following anthesis (Krömer *et al.*, 2012), which could enable pollination by both nocturnal and diurnal visitors (Dar *et al.*, 2006; Ortega-Baes *et al.*, 2011).

In this study, we analysed the floral and reproductive biology of *T. macropetala*. Specifically, (1) the floral visitors that effectively pollinate the flowers were identified, (2) the floral

phenology was described, (3) the production pattern and concentration of nectar were determined, and the floral scent was analysed, and (4) the reproductive system of the bromeliad was assessed. As anthesis in this species begins at dusk and extends into the following day, our primary hypothesis was that *T. macropetala* receives both nocturnal and diurnal visits, but because nocturnal visitors visit the receptive flower first, they will be more important pollinators, accounting for the majority of fruits or seeds. Finally, as suggested by the dilute and hexose-rich nectar, we hypothesized that bats are the main pollinators of this species.

MATERIALS AND METHODS

Study site

Fieldwork was carried out in March and April of 2011 and 2012, in the municipality of Tlalnelhuayocan, in the central region of Veracruz State, Mexico. The site has an elevation of 1500–1700 m (Mehltreter *et al.*, 2005), with an annual precipitation of 1650 mm and an average temperature of 14 °C (Williams-Linera, 2002). The predominant vegetation is humid montane forest (Williams-Linera, 2007). Monitoring of *T. macropetala* phenology was conducted in 2011 in a forest fragment (19°31'12.9"N, 96°59'17.9"W; Fig. 1) where the dominant trees were *Liquidambar macrophylla* Oerst and *Quercus* spp., and

which is surrounded by a mosaic of anthropized vegetation. For logistical reasons, the study in 2012 was conducted in another fragment 1.2 km from the original site (19°31'53.7"N, 96°58'46.8"W; Fig. 1). This second fragment featured similar vegetation, but was surrounded by fruit trees (*Citrus* sp., *Macadamia* sp., *Musa* sp.) and elements of secondary forest.

Study species

Tillandsia macropetala is an epiphytic bromeliad that is generally found on the trunks and lower parts of its host trees but can also be lithophytic or terrestrial (Krömer *et al.*, 2012; Fig. 2A). Its leaves form a stemless tank-type rosette of almost 1.2 m in diameter; its inflorescence is 55–83 cm in length and is composed of 3–7 spikes, each with 15–20 flowers. The flowers (Fig. 2B) are distichous and erect, actinomorphic and with a subsessile helicoiform corolla. The petals are light green, strap-shaped, twisted, and 10.7–12.3 cm long and 14–17 mm wide. The light-green to white stamens are subequal, with free filaments 10.4–11.7 cm long; the anthers are green to green-whitish, curved and 7.5–8 mm long. The style is free and has a trilobulate stigma (Krömer *et al.*, 2012).

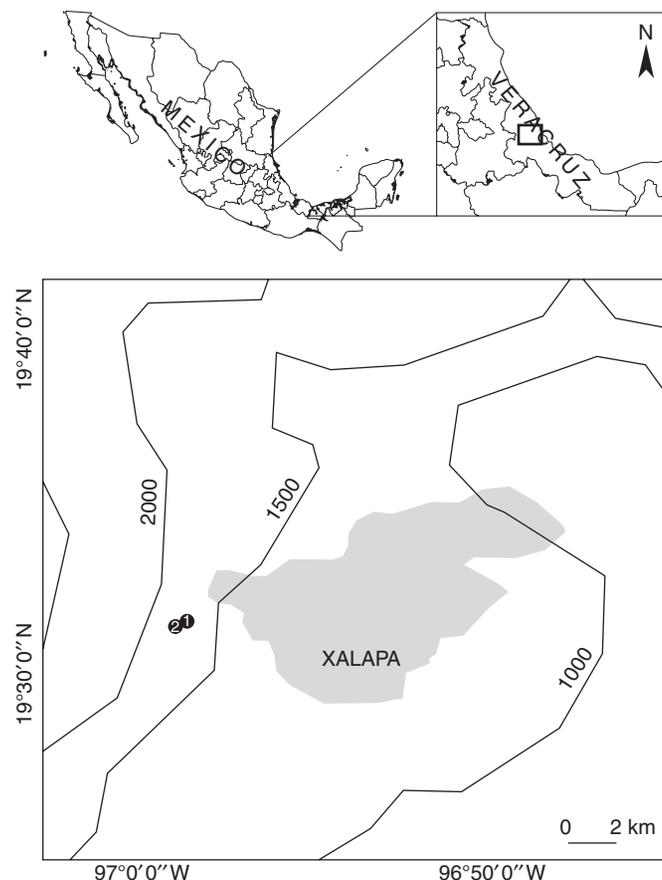


FIG. 1. Study area location in the municipality of Tlalnelhuayocan, in central Veracruz, Mexico. Study sites are marked with numbers 1 (2011) and 2 (2012).

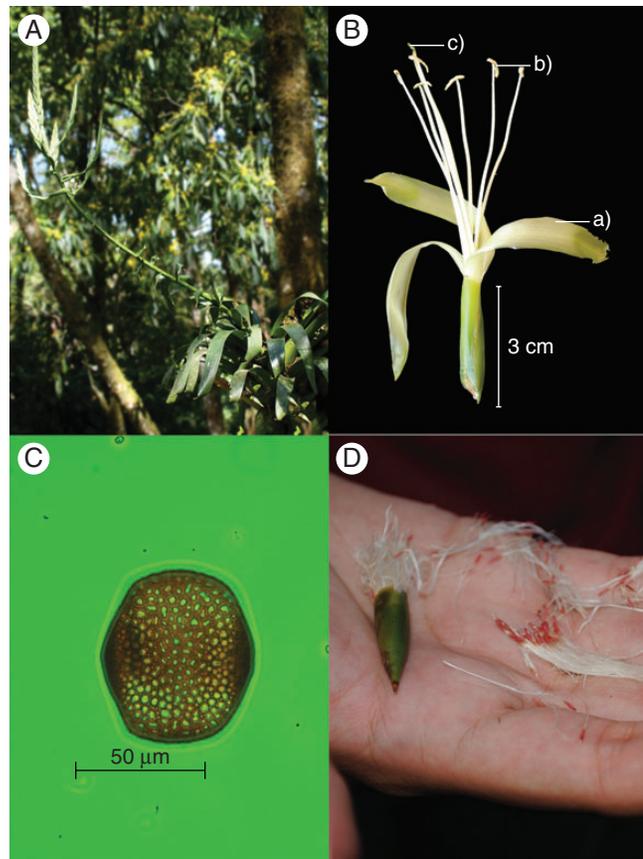


FIG. 2. *Tillandsia macropetala*. (A) Individual with inflorescence in its habitat; (B) flower showing the light green petals (a), as well as the extended stamens (b) and style (c); (C) polar view of a pollen grain (phase contrast technique, 25 \times , Optovar 2); (D) fruit and seeds, the latter with a plumose appendage. Photographs: P. A. Aguilar-Rodríguez, M. C. MacSwiney G. and A. Knauer.

Phenology and floral anthesis

To follow the phenology of *T. macropetala*, 17 individuals that were close to flowering were chosen. Each floral bud was marked with a non-toxic indelible pen on the bract to enable monitoring of the development of individual flowers until fruiting. Once flowering began, floral condition was recorded every day for 3 weeks, describing flower growth, initiation of anthesis, opening of the corolla, and withering of the petals and style (Cascante-Marín *et al.*, 2005; Martín-Rodríguez and Fenster, 2008). Receptivity of the stigma was determined by direct observation of the presence of mucilage on its surface (Escobedo, 2007) and to prove their status, the stigmas of six flowers were dipped in hydrogen peroxide 1, 6, 12, 24 or 36 h after anthesis (Martín-Rodríguez and Fenster, 2008).

In the study population, the majority of *T. macropetala* individuals that were close to flowering were found at heights that impeded manipulation of the flowers (3–5 m). Therefore, nearby plants were translocated and placed at a height of 1.5 m on the trunks of the trees. As some of the individuals of *T. macropetala* used in this study were found at this lower height, it was considered that the translocation should not affect the incidence of floral visitors (as confirmed by preliminary field observations). Regular trips were made to the study site, from December to February, to determine the onset of flowering.

Reproductive system

To determine the reproductive system of *T. macropetala*, pollination treatment experiments were carried out on six individuals placed in pots with gravel in a structure covered with mosquito netting that prevented visits to the flowers (hereafter referred to as the 'planthouse'). The planthouse provided shade and temperatures similar to that found in the trees, and irrigation was carried out sporadically.

In the planthouse, the flowers were subjected to four treatments: (1) emasculation (apomixis) ($n = 25$ flowers) consisted of the emasculation of the flowers by removal of the anthers, to prevent pollen deposition on the flower; (2) in spontaneous self-pollination ($n = 50$ flowers), anthers were not removed and no other manipulation was carried out; (3) cross-pollination (xenogamy) ($n = 40$ flowers), in which the stigma was covered with pollen from the anthers of another plant individual; and (4) self-pollination (autogamy) ($n = 29$ flowers), in which the stigma was covered with pollen from the anthers of the same individual plant. All treatments were applied to previously covered virgin flowers no more than 2 h after the onset of anthesis. Following treatment, the flowers were kept covered to avoid subsequent visits. Eight weeks after conducting the treatments, the presence of fruits was recorded. Percentage of fructification, or fruit-set, was calculated for each treatment and, once the fruits were ripe, the number of seeds per fruit, or seed-set, was recorded (Fig. 2D).

A chi-square test was used to compare percentage of fructification between treatments. To measure the influence of pollination treatments on seed-set (Schmid *et al.*, 2011a), an ANOVA was performed. As the data were not normally distributed (Kolmogorov–Smirnov test), a non-parametric Kruskal–Wallis test with Tukey-type comparisons was used. Analysis was carried out using the program Statistica (ver. 7, StatSoft

Inc., Tulsa, OK, USA) with a level of significance of $P \leq 0.05$. To quantify the self-compatibility of *T. macropetala*, we calculated the self-compatibility index (SC) and the autogamy index (AI) (Wendt *et al.*, 2001; Kamke *et al.*, 2011). The AI was multiplied by 100 to obtain a percentage value of self-compatibility (Martín-Rodríguez and Fenster, 2008).

Nectar analysis

To determine the production pattern and availability of nectar for potential visitors, 21 flowers were randomly chosen from the six individuals kept in the planthouse (comprising at least three flowers from each individual). There was no experimental pollination treatment on any of these flowers, and all measurements of nectar were carried out during periods of no rain. Nectar production was recorded in these flowers every 2 h, using 80- and 10- μ L capillary tubes. Extraction continued until no nectar was left. Total nectar volume per flower was calculated as the sum of the hourly values (Tschapka and von Helversen, 2007). The percentage of sugars contained in each nectar sample was measured using a field refractometer (Mod. HRT32, range: 0–32 %, w/w, precision: 0.2 %; A. Krüss Optronic, Hamburg, Germany). A Pearson correlation was conducted to determine the relationship between volume and concentration of sugars in the nectar and the period during which the measurements were taken.

Floral scent analysis

Scent was collected from two single flowers for about 3 h between 0000 and 0330 h. We used the dynamic pull headspace method for the collection of floral volatiles as described by Huber *et al.* (2005), using Tenax GR (Tenax TR 60/80, Scientific Instrument Services, Old York, RD, USA) as absorbent. We collected scent from an empty oven bag as a control for air contamination.

For chemical analysis of headspace samples, gas chromatography with mass selective detection (GC-MSD) with thermodesorption was used as described by Sun *et al.* (2014). Compound identification was conducted using a mass-selective detector (Agilent MSD 5975, Agilent Technologies, Palo Alto, CA, USA) and ChemStation Enhanced Data Analysis program (version E.02.02). Compounds were identified by comparison of spectra obtained from the samples with those from a reference library (NIST '05 library, <http://www.nist.gov/>). Only floral scent compounds found with larger amounts in the samples collected from flowers than from the control were considered to be emitted by *T. macropetala* flowers.

Observations of floral visitors

Any animal that made contact with the corolla of the flower was considered a floral visitor (*sensu* Schmid *et al.*, 2011b). To be considered potential pollinators, they also had to enter the corolla (Muchhala, 2006) and come into contact with the reproductive organs of the flowers (Schmid *et al.*, 2011b).

Recording of nocturnal visitors was carried out using a video camera (DCR-SR65, Sony Corporation, Tokyo, Japan) in night vision mode, equipped with an infrared light (HVL-HILR, Sony). The camera was placed on a tripod 1.3–1.5 m above the ground and a distance of 1.5 m from the flower. Nocturnal

recording took place from 1900 to 2300 h, which included the initiation of anthesis close to dusk, over a period of 13 d (dictated by a combination of equipment capacity and environmental conditions).

Diurnal visitors were observed directly with binoculars and a digital camera from 0600 to 1100 h (morning period) and from 1500 to 1800 h (evening period), at a distance of around 3 m from the flowers. Direct observations were subsequently made for 1 h before dawn and after dusk, over a period of 16 d. Recordings and observations of visitors were carried out from February to April in both years.

During the observations, the species (or morphospecies) of the visitor was recorded, along with the time of the visit, number of flowers visited and the reward sought. The recordings were analysed using the program Final Cut Pro 7 (Apple Inc. 2009) at a speed of 3 f.p.s. (10 % of real time). For the analysis, the duration of each visit was defined as the amount of time that the mouthparts of the pollinator remained within the corolla (Muchhala, 2006). If a visitor moved the whole inflorescence when placing its head inside the flower, it was considered that contact had been made with the reproductive organs of the flower (Slauson, 2000).

To identify the bat species, sampling was carried out with mist-nets (6 × 12 m) for a period of six nights. The nets were placed at least 1 m from a different *T. macropetala* individual in flower per night, and checked every 30 min. Nets were opened at dusk (1900 h) and closed after 6 h. Species were identified using field guides (Reid, 2009) and following the taxonomy of Simmons (2005) and Velazco and Patterson (2013). Samples of pollen were taken from the bodies of the captured individuals using a moist brush, and placed in vials with 70 % ethanol. Six preparations were made from each sample (18 preparations in total) for subsequent comparison with reference samples of *T. macropetala* pollen grains (Fig. 2C), using an optical microscope (40×, Carl Zeiss, Oberkochen, Germany).

Effectiveness of floral visitors

The effectiveness of the floral visitors was evaluated using floral visitor exclusion treatments (Montalvo and Ackerman, 1986; Sahley, 2001; Wendt *et al.*, 2001). These treatments were conducted on 17 plants in the field, as follows: (1) diurnal exposure (DE) in which the stigmas of the treated flowers ($n = 33$) were covered with a plastic tube (of approx. 3 × 0.5 cm) closed with cotton during the night and uncovered for the diurnal visitors; (2) nocturnal exposure (NE) in which the stigmas of the flowers ($n = 79$) were left exposed during the night and covered during the day; (3) emasculated diurnal exposure (EDE) was carried out as in DE, but in previously emasculated flowers ($n = 33$); (4) emasculated nocturnal exposure (ENE) was carried out as in NE, but in previously emasculated flowers ($n = 62$); and (5) control in which flowers were left continuously exposed to visitors ($n = 136$).

To determine the effectiveness of the visitors, as well as the fruit-set and seed-set produced by the field treatments, we considered the frequency of visits, number of visits in which the visitors pollinated the flower and the behaviour of each visitor during its visit to the flower (e.g. consumption of pollen or petals, form of approach, number of flowers visited and reward sought; Montalvo and Ackerman, 1986).

RESULTS

Phenology and floral anthesis

Flowering of *T. macropetala* began in the third week of December and finished in the second week of April. Opening of the floral bud initiated at approx. 1600 h. The stigma protruded almost 2 h before anthesis and, on reaching its maximum length (12.90 ± 1.51 cm, $n = 12$ flowers), the petals opened to expose the stamens with the dehiscent anthers. Anthesis began around 1900 h (range: 1820–2140 h, $n = 71$ flowers), coinciding with the local time of dusk (approx. 1845–1900 h; Fig. 3). Receptivity of the stigma lasted for around 18 h ($n = 12$ flowers) after opening of the flower.

Senescence began when the floral structures lost colour and turgor between 20 and 24 h after anthesis, completely restricting the entrance of the calyx at around 34 h after the onset of anthesis. In covered flowers without visits, it took about 11 h for the pollen to fall from the anthers ($n = 12$ flowers) by anther senescence. The inflorescence of an individual plant flowered over a period of 36.36 ± 9.51 d ($n = 16$ inflorescences), with an average of 0.7 ± 0.3 flowers d⁻¹ (mean ± s.d.; range: 0–5 flowers daily; $n = 282$ flowers from 14 individuals). The flowering of a spike started at the base and progressed towards the apex over the following days.

Reproductive system

Highest fruit-set was obtained in the self-pollination (34.78 %) and cross-pollination (34.48 %) treatments. There were no fruits formed by the emasculation treatment and fructification was reduced in the spontaneous self-pollination treatment (Table 1). There was a significant difference in the percentage of fructification among treatments ($\chi^2 = 37.169$, d.f. = 4, $P < 0.05$). Excluding the emasculation treatment, which did not produce seeds, there were no significant differences in the number of seeds between the cross-pollination and self-pollination treatments, whereas there was a significant difference between both of these and the spontaneous self-pollination treatment ($H = 5.46$, d.f. = 2, $P < 0.05$; $n = 98$, flowers of six plants; Table 1). According to the SC and AI indices, *T. macropetala* is a self-compatible species (SC = 0.96, AI = 0.34 or 34 %).

Nectar analysis

Mean total volume of nectar of the *T. macropetala* flowers was 434.04 µL (± 178.03 s.d., $n = 21$ flowers in six individuals) per flower per night, and the average concentration of dissolved sugars was 7.43 % (± 6.34 % s.d., $n = 21$ flowers). Nectar was produced for 12 h. The highest quantity of nectar was found at 2200 h (147.02 ± 97.38 µL), subsequently declining to almost zero by 0800 h (1.03 ± 3.90 µL; Fig. 3). The highest concentration of dissolved sugar was recorded at 2000 h, with 15.26 % (± 1.45 %) and this decreased with diminishing nectar production (Fig. 3). Pearson correlation showed that a significant negative relationship existed between the time at which nectar was extracted (conducted at intervals of 2 h) and the volume produced ($r = -0.72$; $P < 0.05$), as well as between the time and the concentration of dissolved sugars ($r = -0.78$; $P < 0.05$).

Floral scent analysis

Nine floral scent compounds were identified in the samples collected from two *T. macropetala* flowers [amounts are given

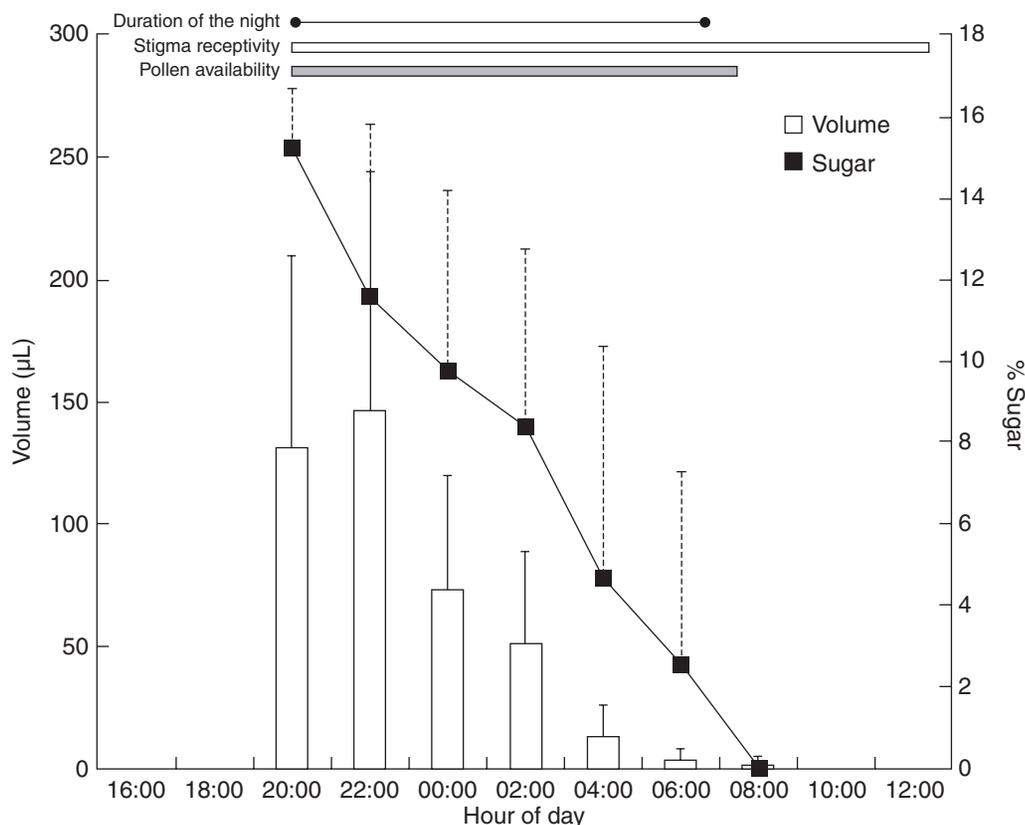


FIG. 3. Production (volume) and concentration (% sugar) of nectar in *Tillandsia macropetala* flowers. Error bars denote s.d. The duration of the receptivity of the stigma and the duration of the night at the study site are indicated, together with the period of dehiscence of the anthers and the time at which pollen grains are found in these.

TABLE 1. Results of the pollination experiments carried out with *Tillandsia macropetala* flowers for determination of the reproductive system

Treatment	No. of flowers manipulated	No. of fruits	Fruit-set	Seed-set (mean \pm s.d.)
Emasculation	24	0	0	0
Spontaneous self-pollination	46	6	13.04	50.6 \pm 138.5 ^a
Cross-pollination	29	10	34.48	154.0 \pm 225.6 ^b
Self-pollination	23	8	34.78	148.0 \pm 215.0 ^b

Different letters denote significant differences ($H = 5.46$, d.f. = 2, $P < 0.05$; the emasculation treatment was excluded).

in per cent of total peak area (%): three fatty acid derivatives (nonanal, 19.8%; heptanal, 10%; 1-octen-3-ol, 2.5%) and six terpenoids (limonene, 24.2%; geraniol, 11.5%; methyl geranate, 11%; β -pinene, 8.3%; 6-methyl-5-hepten-2-one, 7.2%; (Z)- β -ocimene, 5.6%).

Observations of floral visitors

A total of 107 h of observation were carried out (29 h at night and 78 h during the day) at 158 *T. macropetala* flowers (53 at night and 105 during the day) during which 210 visits were

recorded (170 at night and 40 during the day), comprising nine different species of floral visitors (three during the night and six during the day; Supplementary Data Table S1).

Thirty-three individual bats of six species were captured near the flowers: *Anoura geoffroyi* Gray ($n = 3$ individuals), *Artibeus lituratus* Olfers ($n = 2$), *Carollia sowelli* Baker, Solari & Hoffmann ($n = 8$), *Diphylla ecaudata* Spix ($n = 2$), *Myotis volans* Allen ($n = 3$) and *Sturnira hondurensis* Anthony ($n = 15$). All individuals of *A. geoffroyi* had pollen on their snouts, chest, forearms and/or wing membranes, and even on the top of the head (Fig. 4A). All the pollen grains identified in the preparations of pollen taken from the fur of *A. geoffroyi* (Fig. 4A) belonged to bromeliads of the subfamily Tillandsioideae (Halbritter 1992), probably from *T. macropetala*, as it was the only flowering bromeliad with crepuscular anthesis at the time of this study. The video recordings showed that *A. geoffroyi* (identified by its size and reduced uropatagium) was the only bat species visiting *T. macropetala*.

Nocturnal video recordings totaling 29 h were obtained, in which 170 visits by three species were recorded: the bat *A. geoffroyi*, at least two mice *Peromyscus* sp. and a nocturnal moth (Noctuidae; Supplementary Data Table S1). *Anoura geoffroyi* made 158 visits of 0.17 ± 0.08 s duration ($n = 10$ visits). Visits began at dusk and continued throughout the 3-h recording period, peaking between 1930 and 2130 h. In all visits, the bats placed their heads within the flowers and on 152 occasions

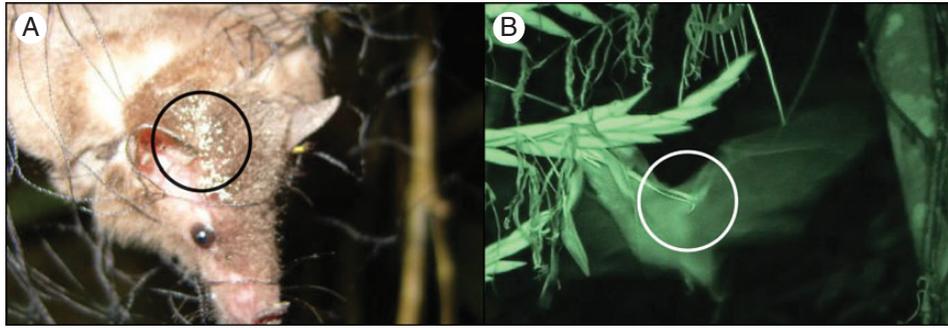


FIG. 4. Images of: (A) Tillandsioideae pollen grains, presumably of *Tillandsia macropetala*, in the fur of an *Anoura geoffroyi* individual captured in a mist-net (circled in black); (B) nectarivorous bat making contact with the anthers of *T. macropetala* during its visit (circled in white). Images: P. A. Aguilar-Rodríguez.

TABLE 2. Results of the exclusion experiments with *Tillandsia macropetala* flowers for determination of the effectiveness of the pollinators

Treatment	No. of flowers	No. of fruits	Fruit-set	Seed-set (mean \pm s.d.)
Diurnal exposure	33	0	0	0
Nocturnal exposure	72	25	34.72	184.5 \pm 268.8 ^a
Emasculated diurnal exposure	31	0	0	0
Emasculated nocturnal exposure	57	22	38.59	199.3 \pm 284.8 ^a
Control	126	49	38.88	180.5 \pm 249.9 ^a

Different letters denote significant differences at $P \leq 0.05$ (diurnal exposure and emasculated diurnal exposure were excluded).

(96.20 %) made contact with the stigma and the stamens, causing the whole spike to move as a result of the force of contact (Fig. 4B). The *Peromyscus* sp. mice were recorded making ten visits in the same night to at least six different flowers in one single bromeliad inflorescence (Aguilar-Rodríguez et al., 2013). The moth was recorded on the *T. macropetala* corolla on two occasions, but was never observed taking the floral rewards.

Forty diurnal visits were recorded. The most frequent diurnal visitor was the hummingbird *Lampornis amethystinus* Swainson, which made 32 visits to the flowers of *T. macropetala*. Of these visits, 28 occurred between 0630 and 0915 h and four between 1800 and 1900 h. As a result of its way of approaching the flowers (not pressing the head against the flower, unlike the bat), in combination with the open corolla, *L. amethystinus* hardly made contact with the reproductive organs of *T. macropetala*.

Effectiveness of floral visitors

Only the flowers exposed to nocturnal visitors (treatments NE and ENE) developed fruits, with both nocturnal exposure treatments achieving 40 % fructification (Table 2), whereas the flowers exposed to diurnal visitors (treatments DE and EDE) did not produce fruits. Seed-set in treatments NE and ENE did not differ significantly from those of the control treatment ($H = 0.17$, d.f. = 2, $P > 0.05$; $n = 255$ flowers of 17 plants, excluding those of treatments DE and EDE; Table 2).

DISCUSSION

Tillandsia macropetala is a self-compatible species and, although anthesis covered both nocturnal and diurnal periods, displays a pollination system that is specialized towards nocturnal visitors, of which the bat *A. geoffroyi* is the only pollinator. Thus, we report for the first time bat-pollination of a species in the genus *Tillandsia*. In bromeliads, chiropterophily has been previously reported for species of the genera *Encholirium*, *Guzmania*, *Pitcairnia*, *Puya*, *Vriesea* and *Werauhia* (Sazima et al., 1989; Krömer et al., 2007; Tschapka and von Helversen, 2007; Fleming et al., 2009; Christianini et al., 2013).

Tillandsia macropetala can be pollinated either by its own pollen or by that of another individual, a common characteristic in bromeliads (Matallana et al., 2010). However, differences in fructification between the spontaneous self-pollination (13 %) and those of the cross-pollination and self-pollination (approx. 35 %) treatments indicate that, while it has the potential for self-pollination, more fruits are developed when an external agent delivers pollen from another plant individual. This may be due to the floral morphology of the species, which makes self-pollination difficult.

Contrary to our hypothesis, even if *T. macropetala* receives nocturnal and diurnal visits, the visitor exclusion treatments conducted in the field showed that only visits made by nocturnal animals produced fruits, thus making the diurnal visitors nectar thieves. Both the duration of stigma receptivity (approx. 18 h, of which 9 h are nocturnal) and the period during which pollen is available on the anthers (approx. 11 h, of which 9 h are nocturnal) make nocturnal pollination more feasible. Nectar production patterns, in terms of volume and concentration, are higher in the initial hours of the night and decrease to a minimum in the initial hours of the morning. In this way, the highest potential reward is available during the night, a trait that has been previously observed in bromeliads visited by bats (Tschapka and von Helversen, 2007).

In accordance with our second hypothesis, the only effective pollinator of *T. macropetala* was the bat *Anoura geoffroyi*, and the captured individuals of this species were found to carry bromeliad pollen of presumably this species. Although *Anoura* bats visit different flowering species, as revealed by different pollen morphotypes (Muchhala and Jarrín-V., 2002), we only found pollen of the same morphotype, which may indicate that night-flowering Tillandsioideae bromeliads were the only source for nectar in the vicinity at the time of this study. It is known that *A. geoffroyi* visits at least two other bromeliad species, namely

Vriesea longiscapa Ule in the coastal tropical lowland forest of south-eastern Brazil (5–90 m; Sazima et al., 1999) and *Vriesea platynema* Gaudichaud at a montane humid forest of Brazil (1100 m; Kaehler et al., 2005). Early morning field observations on the flowers recorded in the previous night showed that in flowers visited by bats, pollen was frequently found on the stigma and the anthers were found without remaining pollen. Due to this lack of pollen available in the morning, diurnal visits probably had no effect on the production of seeds. The predominance of nocturnal visitors to *T. macropetala* is clear from the low frequency of diurnal visitors (19% of the total visits), which act only as nectar thieves (*sensu* Irwin et al., 2010), as no diurnal visitor was observed to make contact with the anthers and/or stigma while taking nectar.

The morphology of the flowers of *T. macropetala* enables many animals (e.g. hummingbirds and various insects) to access the floral reward while making no contact with the reproductive organs. The free extended filaments and style imply that arthropods, because of their small body size, cannot make contact with both anthers and stigma during one visit. In the case of the hummingbird, however, it is the form of approach and visit to the flower that impedes pollination (see Muchhala, 2006).

Considering the characteristics of chiropterophilous plants, the floral morphology of *T. macropetala* does not match the typical traits of large, zygomorphic and bell-shaped flowers with strong scent (Tschapka and Dressler, 2002), such as presented by bromeliads of the genera *Vriesea* and *Werauhia* (Vogel, 1969; Sazima et al., 1995; Cascante-Marín et al., 2005; Krömer et al., 2007). Also, sulphur-containing compounds are absent from the scent of *T. macropetala*, unlike in other bat-pollinated plants (Bestmann et al., 1997; von Helversen et al., 2000). It has also been reported that the nectar of some chiropterophilous bromeliads has a strong and disagreeable odour (e.g. Sazima et al., 1989), whereas the nectar of *T. macropetala* has a faintly sweet odor in the early hours of the night when the volume of nectar is highest (P. A. Aguilar-Rodríguez, pers. obs.).

However, the nocturnal anthesis of *T. macropetala*, its temporal pattern and characteristics of nectar production, extended stamens and stylus, petal colour and phenology, and floral display (annual flowering, with about one flower per night for several nights) match those expected for a bat-pollinated species (Sazima et al., 1999; Tschapka and von Helversen, 2007; Wilmer, 2011).

Nectar sugar concentration of *T. macropetala* is low for a bat-pollinated flower, being about 10% below the average (Tschapka and Dressler, 2002). Comparing nectar characteristics of chiropterophilous bromeliads, the nectar sugar concentration of *T. macropetala* is lower than those of *Guzmania*, *Vriesea* and most *Werauhia* species (Krömer et al., 2008), but the nectar volume is higher ($434.04 \pm 178.03 \mu\text{L}$) than those recorded for *Vriesea longiscapa* and *V. bituminosa* Wawra ($116 - 235 \mu\text{L}$; Sazima et al., 1999) and *T. heterophylla* ($82.21 \pm 48.13 \mu\text{L}$; P. A. Aguilar-Rodríguez, pers. obs.).

The most studied chiropterophilous bromeliad, *Werauhia gladioliflora* (H. Wendland) J.R. Grant, produces more than twice the fruit-set (87 vs. 38%) and ten times as many seeds as *T. macropetala* (1871–2018 vs. 180 seeds; Cascante-Marín et al., 2005; Tschapka and von Helversen, 2007). These differences may be due to the fact that *W. gladioliflora* has a high potential for self-pollination, and may be pollinated by at least four species of bats in the tropical lowland rain forest of Costa Rica

(Tschapka and von Helversen, 2007), whereas *T. macropetala* has only limited capacity for spontaneous self-pollination and was pollinated only by one bat species at our study site. The higher number of bat pollinators may explain the differences in the volume and sugar concentration in nectar of *W. gladioliflora* compared with *T. macropetala* ($1129 \mu\text{L}$ and 17% vs. $434.04 \mu\text{L}$ and 7.43%; Tschapka and von Helversen, 2007).

Unlike the floral scent of *W. gladioliflora*, which included dimethyl disulphide, an unusual component of floral scents that attracts even naïve flower-visiting glossophagine bats (von Helversen et al., 2000), *T. macropetala* scent lacks this compound. However, not all bat-pollinated species have dimethyl disulphide in their scent (Knudsen and Tollsten, 1995), and six out of nine detected compounds in the *T. macropetala* scent have been detected in other bat-pollinated plants (Knudsen and Tollsten, 1995; Bestmann et al., 1997). Also, it is known that plants with an extended floral anthesis change the pattern and components of their floral scent (e.g. *Silene otites*, Caryophyllaceae; Dötterl et al., 2012). For practical reasons, we could only evaluate the floral scent compounds from 0000 to 0330 h from two different flowers, so perhaps monitoring the scent emission through the entire anthesis of *T. macropetala* – including the period with maximum number of visits – would reveal additional volatile compounds.

Our results on the pollination biology of this bromeliad are in accordance with the isolated phylogenetic position of *T. viridiflora* (the type species of *Pseudalcantarea*) as sister to the rest of *Tillandsia/Racinaea/Viridantha* (Barfuss et al., 2005). This might indicate an ongoing evolution from bird or moth pollination, which are common in *Tillandsia* (Benzing, 2000; Kessler and Krömer, 2000), to bat-pollination, in the same way as suggested for the origin of chiropterophily (von Helversen and Winter, 2003). Likewise, the present study supports Krömer et al.'s (2008) suggestion, based on the characteristics of the nectar, that this bromeliad may be considered a chiropterophilous species.

However, note that at our study site *T. macropetala* is at the northern limits of its distribution, and in the more tropical areas of Mexico and in Central America there are more species of nectar-feeding phyllostomid bats (Espinoza et al., 1998; Estrada and Coates-Estrada, 2001; Laval and Rodríguez-H., 2002), and also other animals that could possibly serve as pollinators (e.g. hummingbirds and moths). Thus, more detailed studies of nectar characteristics, pollinators and reproductive biology of the relevant species, especially realized in more diverse tropical zones, are necessary to understand the evolution of pollination syndromes in the genus *Tillandsia* in general and the subgenus *Pseudoalcantarea* in particular.

SUPPLEMENTARY DATA

Supplementary data are available online at www.aob.oxfordjournals.org and consist of Table S1: list of principal floral visitors of *T. macropetala* recorded with video cameras and by direct observation.

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SUPPLEMENTARY DATA

Principal floral visitors of *T. macropetala* recorded with video cameras and by direct observation.

Period of activity	Species	Relative frequency (N) %	Visit reward / objective	Visitor category
Nocturnal	<i>Anoura geoffroyi</i> (Chiroptera: Phyllostomidae)	(158) 75.24	Nectar	Pollinator
Nocturnal	<i>Peromyscus</i> sp. (Rodentia: Cricetidae)	(10) 4.76	Pollen	Pollen thief
Nocturnal	Noctuidae sp.1 (Lepidoptera)	(2) 0.95	-	-
Diurnal	<i>Lampornis amethystinus</i> (Apodiformes: Trochilidae)	(32) 15.24	Nectar	Nectar thief
Diurnal	<i>Dismorphia</i> sp. (Lepidoptera: Pieridae)	(2) 0.95	Nectar	Nectar thief
Diurnal	<i>Bombus</i> sp.1 (Hymenoptera: Apidae)	(2) 0.95	Nectar	Pollen thief
Diurnal	<i>Apis mellifera</i> (Hymenoptera: Apidae)	(2) 0.95	-	-
Diurnal	Vespidae sp.1 (Hymenoptera)	(1) 0.48	Nectar	Nectar thief
Diurnal	Blattidae sp.1 (Blattodea)	(1) 0.48	Nectar	Nectar thief